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The glycine lipids of *Bacteroides thetaiotaomicron* are important for fitness during growth *in vivo and in vitro*

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Abstract
Acylated amino acids function as important components of the cellular membrane in some bacteria. Biosynthesis is initiated by the N-acylation of the amino acid and this is followed by subsequent O-acylation of the acylated molecule resulting in the production of the mature diacylated amino acid lipid. In this study we use both genetics and liquid chromatography-mass spectrometry (LC-MS) to characterize the biosynthesis and function of a diacylated glycine lipid (GL) species produced in Bacteroides thetaiotaomicron. We, and others, have previously reported the identification of a gene, named glsB in this study, that encodes a N-acyltransferase activity responsible for the production of a monoacylated glycine called N-acyl-3-hydroxy-palmitoyl glycine (or commedamide). In all of the Bacteroidales genomes so far sequenced the glsB gene is located immediately downstream from a gene, named glsA, also predicted to encode a protein with acyltransferase activity. We use LC-MS to show that co-expression of glsB and glsA results in the production of GL in Escherichia coli. We constructed a deletion mutant of the glsB gene in B. thetaiotaomicron and we confirm that glsB is required for the production of GL in B. thetaiotaomicron. Moreover, we show that glsB is important for the ability of B. thetaiotaomicron to adapt to stress and colonize the mammalian gut. Therefore, this report describes the genetic requirements for the biosynthesis of GL, a diacylated amino acids species that contributes to fitness in the human gut bacterium, B. thetaiotaomicron.

Importance
The gut microbiome has an important role in both health and disease of the host. The mammalian gut microbiome is often dominated by bacteria from the Bacteroidales, an Order that includes Bacteroides and Prevotella. In this study we have identified an acylated amino acid, called glycine lipid, produced by Bacteroides thetaiotaomicron, a beneficial bacterium originally isolated from the human gut. In addition to identifying the genes required for the production of glycine lipids we show that glycine lipids have an important role during the adaptation of B. thetaiotaomicron to a number of environmental stresses including exposure to either bile or air. We also show that glycine lipids are important for the normal colonization of the murine gut by B. thetaiotaomicron. This work identifies glycine lipids as an important fitness determinant in B. thetaiotaomicron and therefore
increases our understanding of the molecular mechanisms underpinning colonization of the mammalian gut by beneficial bacteria.
Introduction

Members of the Phylum Bacteroidetes, including genera containing important human gut commensal bacteria such as *Bacteroides*, *Parabacteroides* and *Prevotella*, dominate the healthy human gut microbiota (1). The gut-associated Bacteroidetes are required to digest complex dietary glycans into short-chain fatty acids (such as acetate and propionate) that are accessible to the host (2-4). A longitudinal study in infants has revealed the presence of *Bacteroides* in the infant gut within 1 week of birth and some species of *Bacteroides* have been shown to utilize the polysaccharides present in human breast milk (5, 6). Therefore it has been suggested that *Bacteroides* may have an important role during the early development of the infant gut (6).

Acylated amino acids can be found in the membranes of many bacteria (7, 8). The best-characterized, and most widespread, acylated amino acid is ornithine lipid (OL). OL contains a 3'-hydroxy fatty acid group attached by an amide linkage to the \( \alpha \)-amino group of ornithine with a second fatty acid group ester linked to the 3'-hydroxy group of the first fatty acid (9). The genetics of OL production was first described in *Sinorhizobium meliloti* where it was shown that a \( N \)-acyltransferase encoded by *olsB* catalysed the attachment of an hydroxylated fatty acid group to ornithine, resulting in monoacylated ornithine or lyso-OL (10). The second fatty acid was subsequently attached to the hydroxyl group of lyso-OL through the activity of an \( O \)-acyltransferase encoded by *olsA*, resulting in the production of OL (11). An enzyme called OlsF, containing functionally independent domains with \( N \)-acyltransferase and \( O \)-acyltransferase activity, has recently been shown to produce OL in *Serratia proteamaculans* (12). Activities that further modify the OL by hydroxylation or methylation have also been identified (13-15). OL have been shown to be important for growth during acid and temperature stress in *Rhizobium tropici* and depletion of OL results in an increase in the speed of crown gall tumor formation in plants infected with *Agrobacterium* (13, 16). Therefore, OL are important during the interactions between bacteria and their environment, including hosts.

We, and others, have previously identified a gene from *Bacteroides*, initially named choA, that encodes a \( N \)-acyltransferase required for the production of a monoacylated glycine species, called \( N \)-acyl-3-hydroxy-palmitoyl glycine or commendamide (17, 18). We have
previously reported that choA encodes a protein with homology to the N-acyltransferase
domain of OlsF, suggesting a potential role for choA in the production of a diacylated glycine
lipid species (17). In this study we use liquid-chromatography-mass spectrometry (LC/MS) to
show that choA is required for the production of a diacylated glycine lipid (GL) in
Bacteroides thetaiotaomicron and, in line with previous nomenclature, we have renamed
choA to glsB. Using heterologous expression in E. coli we show that GL production also
requires glsA, a gene predicted to encode an O-acyltransferase that is located immediately
upstream from glsB in the B. thetaiotaomicron genome. Finally, we show that glsB is
important for the ability of B. thetaiotaomicron to adapt to stress and colonize the
mammalian gut.

Results

ChoA and ChoB are required to produce diacylated GL in E. coli
We have previously shown that choA, when expressed in E. coli, results in the production of
commendamide, a N-acylated (3-OH C16:0) derivative of glycine with hemolytic activity and
the ability to solubilise cholesterol micelles (17). In all members of the Bacteroidales the
choA gene is located immediately downstream from another gene (nominally called choB)
predicted to encode an O-acyltransferase (17). Together, choA and choB are homologous to
the bifunctional amino acid acyltransferase, OlsF, that is responsible for the production of
OL in S. proteamaculans (see Figure 1). Therefore, we wanted to determine whether choA
and choB might work together to produce diacylated glycine lipids (GL). To do this we
amplified BVU_RSO7720 (choA), BVU_RSO7715 (choB) and choB-choA from B. vulgatus and
cloned the genes into pBAD24 for arabinose-controlled expression in E. coli (resulting in the
formation of pBAD-choA, pBAD-choB and pBAD-choBA, respectively). The cells were
cultured in LB broth and gene expression was induced by the addition of 0.2% (w/v) L-
arabinose to the cultures (as described in Materials and Methods). Cells were harvested and
methanol extracts of the cell pellets were subjected to high-resolution LC/MS analysis. In
cells overexpressing choA, we could detect lipid species that corresponded to a glycine with
an N-acyl substitution of varying carbon chain lengths and degrees of saturation, primarily
14:0, 16:0 (i.e. commendamide), 16:1 and 18:1 (see Figure 2 and Table 1). Notably,
overexpression of choA also resulted in the production of very low levels of diacylated
glycine i.e. 0.7% of the total acylated glycine pool (see Table 1). When cells carrying pBAD-
choBA were analysed we could detect a range of both monoacylated and diacylated glycine species (see Figure 2 and Table 1). The identity of these lipids was confirmed by MS/MS fragmentation although some diacylated glycine species appeared to have mixed fatty acid compositions e.g. in the peak with a m/z value of 510.4 there was a mixture of diacylated glycines substituted with 3-OH-16:0 + 12:0 and 3-OH-14:0 + 14:0 (see Table 1). The total level of monoacylated glycine production in cells overexpressing choBA was approximately 4.5-fold lower than the level observed in cells overexpressing choA alone. Moreover, diacylated glycine production in choBA-expressing cells accounted for 92.7% of the total acylated glycine pool (see Table 1). Importantly, we could not detect monoacylated or diacylated glycine in cells overexpressing choB alone (see Figure 2). Therefore, we propose that, in a mechanism analogous to OL biosynthesis, ChoA N-acylates glycine, resulting in the formation of lyso-GL which is, subsequently, O-acylated by ChoB to produce diacylated GL. In accordance with the nomenclature used for the genes involved in OL biosynthesis, we propose to rename choA and choB as glsA (encoding glycine N-acyltransferase) and glsB (encoding lyso-GL O-acyltransferase), respectively (see Figure 1B).

GlsB is required for the production of commendamide in B. thetaiotaomicron

We wanted to confirm that glsA and glsB were involved in the production of GL in Bacteroides. To do this we decided to take advantage of the genetic tools available in Bacteroides thetaiotaomicron VPI-5482 in order to make a deletion mutant of the glsB (BT_3459) homologue in this bacterium. We also constructed a strain whereby a native copy of glsB was inserted into the genome of the ΔglsB mutant strain (ΔglsB::glsB).

Unfortunately, despite several attempts, we were unable to construct a knock-out mutation of glsA (BT_3458), or a double knock-out of glsA glsB, in B. thetaiotaomicron. Nonetheless, we were able to identify lipid species in both WT and the complemented ΔglsB::glsB strain that were consistent with commendamide and other N-acylated derivatives of glycine (see Figure 3 and Table 2). Further mass spectrometric analysis indicated the presence of diacylated GL in both WT and the complemented ΔglsB::glsB strain but not in the ΔglsB mutant (see Figure 3B and Table 2). These species showed differences in their retention time compared to the GL found in E. coli and gave rise to multiple, partially resolved chromatographic peaks. Bacteroides are known to produce branched-chain fatty acids although from our MS/MS analysis it was not possible to definitively assign whether the
glycines were acylated with straight or branched-chain acyl groups (e.g. 16:0 or methyl-15:0) or if they were iso- or anteiso-branched (19). Importantly, we could not detect any acylated glycine in the ΔglsB mutant (see Table 2). Therefore, glsB is required for the production of all acylated glycine species in *B. thetaiotaomicron*. Interestingly, a comparison of the total lipid chromatogram did reveal both qualitative and quantitative differences between the ΔglsB mutant and both the WT and ΔglsB::glsB strain (see Figure 3A). Although a comprehensive analysis of these differences is not the objective of this study, we did determine that an ion with the same mass as Lipid 654 is produced by *B. thetaiotaomicron* but absent from the ΔglsB mutant (see Figure 3C). Lipid 654 is an acylated serine-glycine dipeptide that has been detected in many members of the Bacteroidetes (20). Lipid 654, also called flavolipin, was first described in members of the *Flavobacterium* and *Cytophaga* (21-23). Some recent studies with *Porphyromonas gingivalis* have implicated Lipid 654 in osteoblast differentiation and atherosclerosis in humans and have also identified Lipid 654 as a potential microbiome-associated biomarker for multiple sclerosis (20, 24-26). In addition, a series of molecules with retention times approximately 14-16 mins and m/z ratios ranging from 1200-1300 were also completely absent from the ΔglsB mutant. The identity of these species is currently under investigation. Therefore, our data confirms that glsB is required for the production of GL and Lipid 654 in *B. thetaiotaomicron* and a mutation in glsB results in significant qualitative and quantitative changes in the lipid profile of the membranes of this bacterium.

**The expression of glsB and glsA is constitutive in B. thetaiotaomicron**

We wanted to examine the expression levels of glsA and glsB in *B. thetaiotaomicron* during growth. Cells were cultured to mid-exponential phase in either rich (BHIS) or defined (DMM) growth media and total RNA was isolated. RT-PCR analysis suggested that glsA and glsB are on different transcripts in *B. thetaiotaomicron* (see Figure 4A). To determine the expression profile of glsA and glsB we examined the data from previously published microarray experiments undertaken using *B. thetaiotaomicron* cultured under different *in vitro* and *in vivo* conditions (27, 28). In TYG broth, a rich growth medium composed of tryptone, yeast extract and glucose, the levels of expression of both glsA and glsB is highest during early exponential phase and the expression of both genes decreases over time (see Figure 4B). A
similar trend is observed during growth in minimal media but the decrease in \textit{glsA} and \textit{glsB} expression over time is not as strong (see Figure 4B). Therefore, the expression of \textit{glsA} and \textit{glsB} may be linked to growth rate. Moreover, \textit{glsA} and \textit{glsB} are also expressed during colonization of the cecum of mice by \textit{B. thetaiotaomicron} (see Figure 4B). Therefore, \textit{glsA} and \textit{glsB} appear to be constitutively expressed in \textit{B. thetaiotaomicron} during growth \textit{in vitro} and \textit{in vivo}.

\textbf{The \textit{glsB} gene is required for normal growth in vitro}

During preliminary experiments we observed that, when colonies were inoculated from agar plates into BHIS broth and incubated overnight, there was significantly reduced growth of the \textit{\Delta glsB} mutant compared to WT cultures. This suggested that \textit{glsB} might be important for the normal growth of \textit{B. thetaiotaomicron}. In order to quantify this observation we set up 10 overnight cultures, from fresh BHIS agar plates inoculated with WT, the \textit{\Delta glsB} mutant or the \textit{\Delta glsB::glsB} strain and the cultures were incubated at 37\(^\circ\)C for 18h at which point the final OD\textsubscript{600} was taken as a measurement of growth. WT and \textit{\Delta glsB::glsB} cultures grown under these conditions reached OD\textsubscript{600} values with a mean of 1.42 +/- 0.042 and 1.25 +/- 0.25, respectively (see Figure 5). However, the OD\textsubscript{600} values of cultures inoculated with the \textit{\Delta glsB} mutant were significantly lower (0.36 +/- 0.23; P<0.001) confirming that the \textit{\Delta glsB} mutant has a strong growth defect under these conditions. Interestingly, when \textit{\Delta glsB} mutant cells from the overnight broth cultures were inoculated into fresh broth cultures there was no observed defect in growth rate between the WT and the \textit{\Delta glsB} mutant (see Figure 5B). Therefore, our data suggests that \textit{glsB} may be required to facilitate adaptation of \textit{B. thetaiotaomicron} to the transition from growth on a solid surface to growth in liquid broth.

\textbf{The \textit{glsB} gene is required for adaptation to stress in vitro}

The transition from growth in solid to liquid media may represent a stress to the bacterium. Therefore, we decided to assess the sensitivity of the \textit{\Delta glsB} mutant to different stresses that would normally be encountered by \textit{B. thetaiotaomicron} i.e. bile stress and the presence of oxygen in air. \textit{B. thetaiotaomicron} was cultured in BHIS broth to mid-exponential phase before the cells were transferred to fresh medium supplemented with 1\% (w/v) porcine bile
and the cells were incubated, anaerobically, for a further 14 h. Under these conditions both WT and ΔglsB::glsB cultures reached a final cell density of $2.6 \times 10^9$ cfu ml$^{-1}$ and $2.7 \times 10^9$ cfu ml$^{-1}$ respectively (see Figure 6A). This is only marginally lower than the cell density achieved when cells are grown under the same conditions but in the absence of bile ($2.98 \times 10^9$ cfu ml$^{-1}$ and $2.9 \times 10^9$ cfu ml$^{-1}$, respectively) and this reflects the high level of bile tolerance associated with the Bacteroides (29). In contrast the ΔglsB mutant only achieved a cell density of $2.7 \times 10^5$ cfu ml$^{-1}$ when cultured in the presence of 1% (w/v) porcine bile (in contrast to $2.7 \times 10^9$ cfu ml$^{-1}$ when cultured in the absence of bile). Therefore, the ΔglsB mutant is approximately $10^4$-fold more sensitive to porcine bile than the WT (see Figure 6A).

Similarly, the ΔglsB mutant exhibited a 10-fold increased sensitivity to exposure to air for 14h compared to both the WT and ΔglsB::glsB strain (see Figure 6B). Therefore, the glsB gene is important in B. thetaiotaomicron to allow adaptation to a variety of stresses including exposure to bile and air.

The glsB gene is required for normal colonization of the murine gut

We wanted to determine the role, if any, of glsB during colonization of the mammalian gut. Therefore, germ-free C57BL/6 mice were subjected to a single, oral gavage of $10^8$ cfu of either WT B. thetaiotaomicron or ΔglsB mutant bacteria. Fecal pellets were collected on Day 2, 6, 9 and 12 post-gavage and bacteria were enumerated by viable plate counting on BHIS agar. On Day 2 the level of WT B. thetaiotaomicron in fecal pellets was $3.6 \times 10^{10}$ cfu g$^{-1}$ feces compared to a significantly lower level of $5.7 \times 10^9$ cfu g$^{-1}$ feces for the ΔglsB mutant (see Figure 7A). However, by Day 4, the ΔglsB mutant was found to be present in fecal pellets at the same level as the WT. Analysis of the cecal contents of mice collected on Day 14 indicated that there is a small, but significant, decrease in the level of the ΔglsB mutant in the cecum compared to WT B. thetaiotaomicron (see Figure 7B). Therefore, the ΔglsB mutant is affected in its ability to colonize the murine gut, particularly during the early stages of colonization.

An important role for B. thetaiotaomicron in the gut is the conversion of dietary glycans into SCFA such as acetate and other organic acids e.g. succinate (4, 30). Therefore, we decided to use acetate and succinate production as a marker of B. thetaiotaomicron metabolism in the
host. Using HPLC, we measured the level of acetate and succinate in the cecal contents collected from germ-free mice infected with either WT or ΔglsB mutant. As expected, we could not detect any acetate or succinate in the cecal contents of uninfected mice, confirming that these metabolites are exclusively derived from microbial activity in the gut (see Figure 7C). There was a small, but significant (Mann-Whitney test; \( P=0.036 \)), decrease in the level of acetate present in the cecal contents of mice colonized with the ΔglsB mutant compared to WT (0.37 mM (0.24-0.53 mM) vs 0.49 mM (0.37-0.94 mM), respectively). The level of succinate was also significantly (Mann-Whitney test; \( P<0.0001 \)) reduced in the cecal contents of mice infected with the ΔglsB mutant compared to the WT bacteria (0.25 mM (0.06-0.38 mM) vs 1.1 mM (0.51-2.3 mM), respectively). Therefore, the metabolism of the ΔglsB mutant is different from WT B. thetaiotaomicron during growth in the murine gut.

Discussion

The acylated amino acids, GL and flavolipin, have previously been identified in the membranes of several different members of the Phylum Bacteroidetes (8, 20, 31). In this study we have identified the genes required for the production of these acylated amino acids. Using genetics and high resolution LC/MS we show that glsB (BT_3459) encodes a glycine \( N \)-acyltransferase that is required for the production of both GL and flavolipin in B. thetaiotaomicron. We also present evidence that glsA (BT_3458), a gene found immediately upstream from glsB on the B. thetaiotaomicron genome, encodes an \( O \)-acyltransferase that is required for the efficient production of GL. Interestingly, the overproduction of glsA and glsB in E. coli results in the synthesis of only GL suggesting the presence of another activity in B. thetaiotaomicron that may convert GL to flavolipin.

Flavolipin (i.e. Lipid 654) has been reported to signal to eukaryotic cells through an interaction with TLR2 (26, 32). The diacylated Lipid 654 has been shown to be converted into a more potent monoacylated derivative, Lipid 430 (or lyso-Lipid 654), through the action of phospholipase A2 activity in the host (20). \( N \)-acyl amino acids are important endogenous signaling molecules produced in the human host e.g. \( N \)-arachidonoyl glycine has been shown to block pain perception in mice (33-35). Genes encoding \( N \)-acyl synthases are enriched in the human gut microbiome and both Lipid 654 and Lipid 430 have been
found in tissues that are distal from the gut indicating that these molecules can be
distributed around the human host (20, 35). Therefore, there is accumulating evidence
supporting an important role for the gut microbiota in the production of acylated amino
acids, such as flavolipin, that can act as signaling molecules in the host.

During periods of phosphate starvation some bacteria access cellular phosphate reserves by
using acylated amino acids such as OL to replace the phospholipids normally found in
bacterial membranes (36-38). On the other hand, OL production in *Rhizobium tropici* and
*Burholderia cepacia* has been shown to be important for the normal tolerance of the
bacterial cell to acid and temperature stress (13, 39, 40). Our data suggests a role for
GL/flavolipin during the response of *B. thetaiotaomicron* to different stresses and a *glsB*
deletion mutant was compromised in its ability to adapt to various stresses including
transition from liquid to solid media, exposure to bile and exposure to air. Interestingly we
could not construct a deletion in *glsA* suggesting that this gene might be essential in *B.
*thetaiotaomicron*. However, IN-Seq analysis of *B. thetaiotaomicron* did not identify either
*glsA* or *glsB* as essential genes although, in the same study, both genes were identified as
being important for normal growth *in vitro* (Goodman et al (2009)). Therefore, it is clear that
*glsA* and *glsB* have an important role in *B. thetaiotaomicron*. Interestingly, the orthologues
of *glsA* and *glsB* have been successfully deleted in *Bacteroides fragilis* suggesting that the
importance of *glsA* and *glsB* during growth might also be species-dependent (41).

In this study we show that the Δ*glsB* mutant in *B. thetaiotaomicron* is affected in its ability
to colonize the gut of a GF mouse. A deletion of both the *glsA* and *glsB* orthologues in *B.
*fragilis* (named *hlyB* and *hlyA*, respectively) was also attenuated for virulence in a mouse
abscess model supporting a role for these genes during in vivo growth (42). We have shown
that, in contrast to the WT, the Δ*glsB* mutant produces decreased levels of both acetate and
succinate whilst in the cecum indicating that there are differences in the metabolism
between the WT and the Δ*glsB* mutant. Acetate and succinate are important end-products
of carbohydrate metabolism in *B. thetaiotaomicron*. Acetate is produced from acetyl-CoA
via the acetate kinase (AckA)-phosphate acetyltransferase (Pta) pathway resulting in the
generation of ATP. The reduction in acetate production in the Δ*glsB* mutant is small and
Succinate production is via phosphoenolpyruvate, oxaloacetate, malate and fumarate and is linked to the production of reducing equivalents (through the regeneration of NAD(P)+) and the formation of a proton motive force.

Fumarate reductase (Frd) catalyzes the production of succinate from fumarate and this protein complex is localized to the inner membrane of Bacteroides. Therefore, one possible explanation for the reduced level of succinate observed in the glsB mutant of B. thetaiotaomicron is that the activity of Frd is compromised by the altered membrane composition of this mutant. Nonetheless, the significantly reduced levels of succinate produced by the ΔglsB mutant would be expected to compromise metabolic flux and, therefore, reduce fitness in the gut. In support of this, both ΔglsA (BT_3458) and ΔglsB (BT_3459) have been identified as important determinants for the colonization of the GF mouse gut during an IN-Seq screen with B. thetaiotaomicron (43). Therefore, we have shown that glsB, and presumably the production of GL and/or flavolipin, is an important fitness factor in Bacteroides, required for the adaptation to stress and normal colonization of the mammalian gut, particularly in the presence of a competing microbiota.

**Materials and Methods**

**Strains, plasmids, primers and growth conditions.**

*Bacteroides thetaiotaomicron* VPI 5482 was cultured anaerobically at 37°C in Brain Heart Infusion media (Sigma), supplemented with hemin (5μg ml-1), 0.1% (w/v) cysteine, and 0.2% (w/v) sodium bicarbonate. *Escherichia coli* EPI300 (Epicentre) was routinely cultured in LB broth at 37°C (Merck). For agar plates 1.5% (w/v) agar was added to the liquid media. Where appropriate, antibiotics were added to the media at the following concentrations: Ampicillin (Amp), 100 μg/ml; Chloramphenicol (Cm), 12.5 μg/ml; Gentamycin (Gm), 50 μg/ml or 200 μg/ml; Erythromycin (Ery), 25 μg/ml. Plasmids and primers used in this study are shown in Table 3 and 4.

**Construction of gene knock-outs in B. thetaiotaomicron**

Gene deletions were carried out using *B. thetaiotaomicron* Δtdk, as previously described (44). Briefly the DNA regions flanking the gene to be deleted were amplified and fused by PCR, cloned into the pEXCHANGE-tdk vector and transformed into *E. coli* S17-1 λpir. The donor (*E. coli*) and recipient (*B. thetaiotaomicron*) strains were mixed, plated onto BHIS agar containing...
10% (v/v) horse blood (BHIS blood agar), and incubated, aerobically at 37°C for 24 h. The biomass was re-suspended in 5ml TYG broth, before plating onto BHIS blood agar supplemented with Gm and Ery. The plates were incubated anaerobically for 48 h at 37°C, before 5-10 colonies were re-streaked onto BHIS blood agar (Gm, Ery). After 48 h at 37°C, single colonies were picked into TYG broth and grown for 20 h without antibiotics before plating onto BHIS blood agar supplemented with 200 μg ml\(^{-1}\) 5-Fluoro-2'-deoxyuridine (FUdR) for vector counter selection. The plates were incubated anaerobically for 72 h and re-streaked onto BHIS blood + FUdR agar plates. Colony PCR, using primers that were designed outside of the flanking regions, was used to identify potential knock out mutants, before confirmation by sequencing. For complementation experiments the glsB (choA) gene, plus 500bp upstream from the proposed translation start site, was amplified, and cloned into the pNbu2-bla-ermGb insertion vector. The cloned DNA fragment was inserted into the \textit{B. thetaiotaomicron Δtdk ΔglsB} genome into either of the two Nbu2-targetted tRNA\text{Ser} loci, via conjugation from \textit{E. coli S17-1 λpir}. The resulting complemented strain, \textit{B. thetaiotaomicron Δtdk ΔglsB::glsB}, was selected by plating onto BHIS blood agar supplemented with Gm and Ery, and the presence of the \textit{glsB} gene was confirmed by PCR.

\textit{Colonization of germ-free C57BL/6NTac mice}

All experiments involving animals were performed at the Biological Services Unit in University College Cork and were approved by the University College Cork Animal Experimentation Ethics Committee. For colonization experiments, 6 week old germ-free female C57BL/6NTac mice were gavaged with 20 μl of 5x10^9 cfu ml\(^{-1}\) of the appropriate bacterial strain (n=9 for \textit{B. thetaiotaomicron} WT, n=8 for \textit{B. thetaiotaomicron ΔglsB} and n=4 for uninoculated control). The mice were housed as groups of 2-3 in individually-ventilated cages (IVC) and bacterial enumerations were carried out by serial dilution and plating homogenized fecal pellets collected from each IVC on Day 2, 6, 9, and 12 post-gavage. All mice were euthanized on Day 14, the ceca were harvested and cecal contents were collected for further analysis, including bacterial enumeration.

\textit{Analysis of short-chain fatty acids (SCFA)}

The level of SCFA in the cecal contents was determined by HPLC using a protocol described previously (45). Cecal contents were weighed and re-suspended in sterile MilliQ water (1:10...
(w/v)) containing several 3-4 mm sterile glass beads (Sigma). The samples were vortexed for 1 min and homogenates were centrifuged at 10,000 x g for 10 min. The supernatants were filter sterilised using a 0.22 μm filter and analysed using HPLC with a refractive index detector (Agilent 1200 HPLC system). A REZEX 8μ% H, Organic Acid Column 300 x 7.8 mM (Phenomenex, USA) was used with 0.01 N H₂SO₄ as the elution fluid, at a flow rate of 0.6 ml min⁻¹. The temperature of the column was maintained at 65°C and 20 μl of each sample was injected for analysis. End-product peaks were identified by comparison of their retention times with those of pure compounds and concentrations were determined from standards of known concentrations.

Identification and quantification of glycine lipids

Overnight cultures of E. coli, with the appropriate plasmids, or B. thetaiotaomicron were inoculated into fresh medium (LB broth with 0.2% (w/v) L-arabinose for E. coli or BHIS broth for B. thetaiotaomicron) at an OD₆₀₀=0.05 and allowed to grow at 37°C until OD₆₀₀=0.5-0.6. At this point 1 ml samples were centrifuged (5 min, 12,000 x g) and the pellets were re-suspended in HPLC grade methanol (Sigma) and 500 pmol N-arachidonyl glycine (NAGly 20:4) (Cayman Chemicals, Ann Arbour, MI, USA) was added as an internal standard. Ethyl acetate was added and the mixture was left at 4°C for 30 min before being centrifuged at 2000 x g for 5 min to remove denatured proteins. The supernatant was collected, evaporated to dryness under nitrogen gas and reconstituted in methanol containing 5 mM ammonium formate (Sigma). LC-MS analyses were performed using a Thermo Exactive Orbitrap mass spectrometer (Thermo Scientific, Hemel Hempsted, UK) equipped with a heated electrospray ionization (HESI) probe and coupled to a Thermo Accela 1250 ultra-high pressure liquid chromatography (UHPLC) system. Samples were injected on to a Thermo Hypersil Gold C18 column (2.1 mm x 100 mm, 1.9 μm) maintained at 50°C. Mobile phase A consisted of water containing 10 mM ammonium formate and 0.1% (v/v) formic acid. Mobile phase B consisted of 90:10 isopropanol/acetonitrile containing 10 mM ammonium formate and 0.1% (v/v) formic acid. The initial conditions for analysis were 65%A/35%B and the percentage of mobile phase B was increased from 35%-65% over 4 min, followed by 65%-100% over 15 min held for 2 min before re-equilibration to the starting conditions over 6 min. The flow rate was 400 μl/min. Samples were analysed in negative ion mode over the mass to charge ratio (m/z) range 250-2000 at a resolution of 100,000. The signals
corresponding to the accurate m/z values for [M-H]-ions of glycine lipid molecular species were extracted from raw LC-MS data sets with the mass error set to 5 ppm. Quantification was achieved by relating the peak area of the glycine lipid species to the peak area of the NAGly 20:4 ISTD. Tandem mass spectrometry (MS/MS) was employed to confirm the identity of glycine lipid species. Samples were infused at a rate of 5 µl/min into a Thermo LTQ-Orbitrap XL mass spectrometer and subjected to higher-energy collision dissociation (HCD) in the Orbitrap analyser. Additional MS³ analyses were performed through collision induced dissociation (CID) in the ion trap. Collision energies ranged from 40-65% and helium was used as the collision gas.

Reverse Transcription PCR (RT-PCR)

Overnight Bacteroides cultures were sub-cultured into fresh BHIS broth to an OD₆₀₀=0.05 and incubated anaerobically at 37°C until the cultures reached mid exponential phase (OD₆₀₀=0.3-0.5). At this stage a 5 ml aliquot was removed, centrifuged and the cell pellet re-suspended in RNAprotect (Qiagen). RNA extractions were carried out using High Pure RNA Isolation Kit (Roche), according to manufacturer’s instructions. For the qualitative determination of gene expression, RNA was reverse transcribed into cDNA using QuantiTect Reverse Transcription Kit (Qiagen), according to manufacturer’s instructions. This cDNA was subsequently used as a template for PCR using 0.5 µl DNA template, 2.5 µl CoralLoad PCR buffer (Qiagen), 100 pmol of the appropriate primers (see Supplementary Table 2), 0.2 mM dNTP mix (Promega), 0.125 µl Taq DNA polymerase (Qiagen), and sterile MilliQ dH₂O, in a final volume of 25 µl. The following PCR conditions were used: 95°C for 5 min (initial denaturation), followed by 35 cycles of 95°C for 30 s, primer specific annealing temperature for 30 s, and 72°C for template specific length of time (1 min per 1kbp DNA). This was followed by a final extension of 72°C for 10 min.

Stress assays

Overnight cultures of the appropriate B. thetaiotaomicron strains were adjusted to OD₆₀₀=0.05 in fresh BHIS broth and incubated for 7h at 37°C anaerobically. At this point the OD₆₀₀ was adjusted to OD₆₀₀=0.2 and each culture was split into 3 equal aliquots whereby one aliquot was cultured anaerobically, another aliquot was exposed to air and the final aliquot
was incubated in BHIS broth with 1% (w/v) porcine bile (Sigma). Following incubation for 14h under the appropriate stress conditions, viable cells were enumerated by serial dilutions and plating onto BHIS agar.

Statistical analysis

All statistical analysis was performed using GraphPad Prism 6.0e for Mac software. All experiments were carried out using biological triplicate samples, unless stated otherwise. The Student’s t-test or Mann-Whitney test were used to compare two different groups of data, as indicated. One-way ANOVA, with Tuckey’s post-test, was used to compare three or more groups of data and differences were considered to be significant if P<0.05.

Acknowledgements

This work was funded by an Investigator Award from Science Foundation Ireland (SFI) to D.J.C. 912/IP/1493) and by funding received through APC Microbiome Ireland, a research institute supported by SFI (SFI/12/RC/2273). S.R.T., M.K.D. and P.D.W. gratefully acknowledge the financial support of the European Regional Development Fund, Scottish Funding Council and Highlands and Islands Enterprise.

Author contributions

D.J.C. conceived the study and A.L. carried out all of the experiments except the lipidomics. P.D.W., S.R.T and M.K.D. carried out all lipidomic experiments and analyses. A.L. and D.J.C. analysed the data and D.J.C. wrote the manuscript with the help of A.L. and P.D.W. All authors reviewed the manuscript.

Bibliography


Figure legends

Figure 1. (A) The protein encoded by the choB gene in Bacteroides vulgatus has predicted homology (30.2% identity over 273 amino acids) with the N-terminus of OlsF (Spro_2569), carrying the O-acyltransferase activity required for the biosynthesis of ornithine lipids (OL). Similarly, the choA gene is predicted to encode a protein with homology (32.6% identity over 264 amino acids) to the C-terminus of OlsF, carrying the N-acyltransferase activity involved in OL biosynthesis. (B) Predicted pathway for the production of glycine lipid (GL) in Bacteroides based on homology with OL production in Serratia proteamaculans. In this schematic the N- and O-acylation involves 3-OH-palmitate and palmitate, respectively.

Figure 2. Identification of glycine lipids in E. coli overexpressing choA and choB. E. coli containing pBAD24, pBAD-choA, pBAD-choB or pBAD-choBA were cultured in the presence of 0.2% L-arabinose until mid-exponential phase and extracted lipids were analysed by LC-MS, as described in Materials and Methods. All samples were spiked with 500pmol arachidonyl (20:4) glycine as an internal standard (Rf=4.98 min, m/z 360.25). (A) The base
peak intensity (BPI) chromatogram showing the range of lipids present in the *E. coli*
membrane. All lipid profiles appear to be qualitatively similar, with the exception of pBAD-
*choA* which shows increased peaks eluting with Rf of approx. 4 min where mono-acylated
glycine molecules would be expected to be eluted. (B) Extracted ion chromatograms of
peaks eluting with Rf values corresponding to mono- or di-acylated glycine species (for
clarity only 3-OH 16:0 (Rf=4.56 min m/z 328.25) and 3-OH 16:0+16:0 (Rf=10.09 min m/z
566.48) are shown. The full list of identified molecules can be found in Table 1). (C) MS/MS
fragmentation of the molecule eluting with Rf=4.56 confirming its identification as 3-OH
16:0-glycine. The same peak in both pBAD-*choA* and pBAD-*choBA* gave identical
fragmentation profiles. (D) MS/MS fragmentation confirming the O-acylation of 3-OH 16:0
resulting in a structure of the m/z 566.48 compound that is consistent with a diacylated
glycine (in this case 3-OH 16:0+16:0). All diacylated glycine molecules detected are listed in
Table 1.

**Figure 3.** Identification of glycine lipids in *B. thetaiotaomicron*. Cells (as indicated) were
cultured in BHIS broth until mid-exponential phase and extracted lipids were analysed by LC-
MS, as described in Materials and Methods. All samples were spiked with 500pmol
arachidonyl (20:4) glycine as an internal standard (Rf=4.98 min, m/z 360.25). (A) The base
peak intensity (BPI) chromatogram showing the range of lipids present in the membrane of
*B. thetaiotaomicron*. Clear qualitative (indicated by arrow) and quantitative differences are
observed between the profiles of WT, Δ*glsB* and Δ*glsB::glsB* strains. (B) Extracted ion
chromatograms of peaks eluting with Rf values corresponding to mono- or di-acylated
glycine species. For clarity the internal standard (arachidonoyl (20:4) glycine; Rf=4.98-5.00
min m/z 360.25) and 2 ions corresponding to 3-OH 16:0 (Rf=4.56 min m/z 328.25) and C32:0
(Rf=9.8 min m/z 566.4) are shown. The full list of identified molecules can be found in Table
2. (C) Extracted ion chromatograms of peak eluting with Rf values corresponding to
flavolipin (lipid 654). The internal standard and the ion corresponding to C32:0 flavolipin
(Rf=9.32 min m/z 653.51) are shown.

**Figure 4.** Analysis of the expression of *glsA* and *glsB* in *B. thetaiotaomicron*. (A) RT-PCR
transcript analysis of expression from the *glsAB* locus. B. thetaiotaomicron cells were grown
to mid-exponential phase in BHIS broth and RNA was extracted and back-transcribed into
cDNA (as described in Materials and Methods). Transcript analysis was undertaken using primers combinations that amplify a region specific to *glsA* (A), *glsB* (B) or the intergenic region (AB). (B) Expression of *glsA* and *glsB* under different *in vitro* and *in vivo* growth conditions. Normalized microarray data was extracted from datasets available in the GEO database (Accession number: GSE2231). Sample preparation and analysis is described in (27, 28). For the *in vitro* samples, *B. thetaiotaomicron* was cultured in chemostats using different media (TYG broth, Minimal Medium-Maltose (MM-M), Minimal Medium-Glucose (MM-G)). At the indicated times cells were harvested, RNA was extracted and expression profiling was undertaken using custom *B. thetaiotaomicron* GeneChips. The data presented are the mean of 2 biological replicates and the error bars represent the standard deviation. For the *in vivo* experiments, individual germ-free NRBI mice (n=9) were monoassociated with *B. thetaiotaomicron* and fed a standard chow diet for 10 days. RNA was extracted from the cecal contents of each mouse and used for expression profiling using the custom *B. thetaiotaomicron* GeneChips.

**Figure 5.** The ∆*glsB* mutant is unable to normally transition from solid to liquid growth media. (A) *B. thetaiotaomicron* (WT, ∆*glsB* and ∆*glsB::glsB*) was grown on BHIS agar and individual colonies (n=10) were inoculated into fresh BHIS broth (solid → liquid). The cultures were incubated at 37°C for 24h and OD₆₀₀ was used to measure growth. Each point represents an individual culture and the mean (+/− standard deviation) is presented (***P<0.0001 as determined using one-way ANOVA with Tukeys post-test for multiple comparisons). (B) Cells cultured in (A) were used to inoculate fresh BHIS broth and cells were grown at 37°C and OD₆₀₀ was measured at the indicated time. Each strain was grown in triplicate and each point is the mean of the replicates and errors bars represent the standard deviation.

**Figure 6.** The *glsB* gene is required for adaptation to stress. *B. thetaiotaomicron* (WT, ∆*glsB* and ∆*glsB::glsB*) was cultured to mid-exponential phase (OD₆₀₀=0.2) before (i) inoculation into BHIS broth followed by anaerobic incubation for 14h (Control); (ii) inoculation into BHIS broth supplemented with 1% (w/v) porcine bile followed by anaerobic incubation for 14h (1% bile); (iii) inoculation into BHIS broth followed by aerobic incubation (with vigorous shaking) for 14h (Aerotolerance). Viable cell counts were determined using serial dilutions
that were plated onto BHIS agar followed by anaerobic incubation for 24-48h. The experiment was repeated 3 times and the error bars represent the standard deviation. Significance was determined using a one-way ANOVA with Tukeys post-test for multiple comparisons (**P<0.01, ***P<0.001, ****P<0.0001).

Figure 7. *B. thetaiotaomicron* requires the *glsB* gene for normal colonization of the gut of mice. Germ-free C57BL/6 mice were gavaged with a single $10^8$ cfu dose of *B. thetaiotaomicron* WT (n=9) or Δ*glsB* mutant (n=8), as indicated. Control mice (n=4) were not colonized. (A) At the indicated time post-gavage fecal pellets were collected and bacteria were enumerated by plating fecal homogenates on BHIS agar. Bacteria were also enumerated in the cecum of colonized mice on Day 14 post-gavage. Control mice did not contain any bacteria at this point. Error bars represent the standard deviation of colonization levels in at least 5 mice (Unpaired t-test *P<0.05, ****P<0.001). (B) The contents of the cecum from each colonized and control mouse was examined for the presence of acetate and succinate by HPLC analysis. The error bars represent the 25-75% percentile values from the median and significance was determined using the Mann-Whitney test (*P<0.05, ****P<0.0001).
Table 1. Glycine lipids in *E. coli* expressing *glsB* and *glsA*

<table>
<thead>
<tr>
<th>Predicted size of acyl group (m/z)</th>
<th>Concentration (pmol/10⁹ cells)±S.D.</th>
<th>pBAD-choA/<em>glsB</em></th>
<th>pBAD-choBA/<em>glsAB</em></th>
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</thead>
<tbody>
<tr>
<td>3-OH-12:0</td>
<td>nd</td>
<td>35.3±9.0</td>
<td>nd</td>
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<tr>
<td>3-OH-14:0 (300.2)*</td>
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<td>221.3±145.2</td>
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<td>3-OH-14:1 (298.2)*</td>
<td>nd</td>
<td>118.0±13.3</td>
<td>nd</td>
</tr>
<tr>
<td>3-OH-15:0 (314.2)*</td>
<td>nd</td>
<td>52.4±14.4</td>
<td>9.2</td>
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<tr>
<td>3-OH-16:0 (328.2)*</td>
<td>nd</td>
<td>3954.6±215.3</td>
<td>1584.9±843.4</td>
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<tr>
<td>3-OH-16:1 (326.2)*</td>
<td>nd</td>
<td>6831.9±264.0</td>
<td>382.3±267.1</td>
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<td>3-OH-18:0 (356.1)*</td>
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<td>Total monoacylated glycine</td>
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<td>3-OH-16:0+14:0 (538.4)*</td>
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<tr>
<td>3-OH-16:1+14:0/3-OH-16:0+14:1 (536.4)*</td>
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<td>3-OH-18:1+16:1 (590.4)*</td>
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<td>3-OH-36:2</td>
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<td>Total diacylated glycine</td>
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<tr>
<td>Total acylated pool</td>
<td><strong>17737.4</strong></td>
<td><strong>53765.0</strong></td>
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</tbody>
</table>

*: not detected in all biological replicates

*: acylated glycine confirmed by MS/MS fragmentation. Acyl group designation is indicative, based on the predicted number of carbons in the lipid species.
Table 2. Glycine lipids in *B. thetaiotaomicron*

<table>
<thead>
<tr>
<th>Predicted size of acyl group (m/z)</th>
<th>Concentration (pmol/10⁹ cells) ±S.D.</th>
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<tbody>
<tr>
<td></td>
<td>WT</td>
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<td>3-OH-15:0/3-OH-methyl 14:0 (314.2)*</td>
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<td>3-OH-16:0/3-OH-methyl 15:0 (328.2)*</td>
<td>252.1±36.3#</td>
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<td>3-OH-17:0/3-OH-methyl 16:0 (342.2)*</td>
<td>429.9±71.6#</td>
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<td>Total monoacylated glycine</td>
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<td>3-OH-methyl 15:0+13:0/3-OH-methyl 14:0+14:0 (524.4)*</td>
<td>223.2±25.5</td>
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<td>3-OH-methyl 15:0+15:0/3-OH-methyl 16:0+14:0 (552.4)*</td>
<td>2000.3±139.3</td>
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<td>3-OH-methyl 16:0+15:0/3-OH-methyl 15:0+16:0 (566.4)*</td>
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<tr>
<td>Total diacylated glycine</td>
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<td>Total acylated pool</td>
<td><strong>6368.1</strong></td>
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- nd: not detected
- #: only detected in 2 (out of 3) biological replicates
- *: acylated glycine confirmed by MS/MS fragmentation. Acyl group designation is indicative, based on the predicted number of carbons in the lipid species.
Table 3. Strains and plasmids used in this study

### Strains

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<th>Characteristics</th>
<th>Source</th>
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<td><em>Escherichia coli</em></td>
<td>F- mcrA Δ(mrr-hsdRMS-mcrBC) Φ80d lacZΔM15 ΔlacX74 recA1 endA1 araD139 Δ(ara, leu)7697 galU galK λ-rpsL (StrR) nupG trfA dhfr</td>
<td>Epicentre</td>
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<td>B. thetaiotaomicron</td>
<td>VPI 5482 with Δtdk deletion; Gm&lt;sup&gt;R&lt;/sup&gt;, F UdR&lt;sup&gt;R&lt;/sup&gt;</td>
<td>Eric Martens, University of Michigan</td>
</tr>
<tr>
<td>B. thetaiotaomicron ΔglsB</td>
<td>Derivative of Δtdk strain with glsB deletion; Gm&lt;sup&gt;R&lt;/sup&gt;, F UdR&lt;sup&gt;R&lt;/sup&gt;</td>
<td>This study</td>
</tr>
<tr>
<td>B. thetaiotaomicron ΔglsB::glsB</td>
<td>ΔglsB strain with complementing glsB + 500bp upstream sequence; Gm&lt;sup&gt;R&lt;/sup&gt;, Erm&lt;sup&gt;R&lt;/sup&gt;</td>
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### Plasmid

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<td>pBAD24</td>
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<td>This study</td>
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<td>This study</td>
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<td>(44)</td>
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<td>pEXCHANGE-Δtdk ΔglsB</td>
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<td>pNbu2-bla-ermGb</td>
<td>inserts into NBU2 att1 and/or att2 sites, Erm&lt;sup&gt;R&lt;/sup&gt;, Amp&lt;sup&gt;R&lt;/sup&gt;</td>
<td>(44)</td>
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<td>pNbu2-bla-ermGb-pglsB</td>
<td>chromosomal insertion vector; inserts into NBU2 att1 and/or att2 sites, Erm&lt;sup&gt;R&lt;/sup&gt;, Amp&lt;sup&gt;R&lt;/sup&gt;</td>
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Table 4. Primers used in this study

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<td>RTfwB300thet</td>
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<td>FwBA630thet</td>
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<td>RT-PCR for glsAB fragment from B. thetaiotaomicron</td>
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<td>FwchoAFLANK1</td>
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<td>glsB deletion from B. thetaiotaomicron</td>
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<tr>
<td>RvchoAFLANK1</td>
<td>GCACTCGATCTTTACCGGAAATTACATATTCTTGTTATAGTGGTTCATTC</td>
<td>glsB deletion from B. thetaiotaomicron</td>
</tr>
<tr>
<td>FwchoAFLANK2</td>
<td>GATAGAACACTATAAACAAGAATATGTAATTTCCGTAAAGATCGAGTGC</td>
<td>glsB deletion from B. thetaiotaomicron</td>
</tr>
<tr>
<td>RvchoAFLANK2</td>
<td>CTGCTGTCTAGATACCCCTTTTCATCGAGGCC</td>
<td>glsB deletion from B. thetaiotaomicron</td>
</tr>
<tr>
<td>FwchoAKOcheck</td>
<td>GGGTTCTATCTGAAGAAAATAG</td>
<td>Checking glsB deletion in B. thetaiotaomicron</td>
</tr>
<tr>
<td>RvchoAKOcheck</td>
<td>TCAACGCTTGCCCTCCATCG</td>
<td>Checking glsB deletion in B. thetaiotaomicron</td>
</tr>
</tbody>
</table>
A)

Bacteroides vulgatus

choB/glsA

BVU_RSO7715

O-acyltransferase

choA/glsB

BVU_RSO7720

Serratia proteamaculans

olsF

B)

Glycine + 3-OH-palmitate

choA/glsB

Lyso-GL

choB/glsA

GL
A) Glycine lipid

B) Flavolipin

C) Glycine lipid

WT

ΔglsB

ΔglsB:ΔglsB

Relative Abundance %

Time (min)