

UHI Research Database pdf download summary

Identification of C25 highly branched isoprenoid (HBI) alkenes in diatoms of the genus *Rhizosolenia* in polar and sub-polar marine phytoplankton.

Belt, Simon T.; Brown, Thomas A.; Smik, Lukas; Tatarek, Agnieszka; Wiktor, Józef; Stowasser, Gabriele; Assmy, Philipp; Allen, Claire S.; Husum, Katrine

Published in:
Organic Geochemistry

Publication date:
2017

Publisher rights:
Copyright © 2017 Elsevier B.V. or its licensors or contributors.

The re-use license for this item is:
CC BY-NC-ND

The Document Version you have downloaded here is:
Peer reviewed version

The final published version is available direct from the publisher website at:
[10.1016/j.orggeochem.2017.05.007](https://doi.org/10.1016/j.orggeochem.2017.05.007)

[Link to author version on UHI Research Database](#)

Citation for published version (APA):

Belt, S. T., Brown, T. A., Smik, L., Tatarek, A., Wiktor, J., Stowasser, G., Assmy, P., Allen, C. S., & Husum, K. (2017). Identification of C25 highly branched isoprenoid (HBI) alkenes in diatoms of the genus *Rhizosolenia* in polar and sub-polar marine phytoplankton. *Organic Geochemistry*, 110, 65-72.
<https://doi.org/10.1016/j.orggeochem.2017.05.007>

General rights

Copyright and moral rights for the publications made accessible in the UHI Research Database are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights:

- 1) Users may download and print one copy of any publication from the UHI Research Database for the purpose of private study or research.
- 2) You may not further distribute the material or use it for any profit-making activity or commercial gain
- 3) You may freely distribute the URL identifying the publication in the UHI Research Database

Take down policy

If you believe that this document breaches copyright please contact us at RO@uhi.ac.uk providing details; we will remove access to the work immediately and investigate your claim.

© 2017. This manuscript version is made available under the CC-BY-NC-ND 4.0 license <http://creativecommons.org/licenses/by-nc-nd/4.0/>

This is a pre-copyedited, author-produced version of an article accepted for publication in Organic Geochemistry following peer review.

The version of record [Belt S.T., Brown T.A., Smik L, Tatarek A, Wiktor J, Stowasser G, Assmy P, C, A. and K, H. (2017) Identification of C25 highly branched isoprenoid (HBI) alkenes in diatoms of the genus Rhizosolenia in polar and sub-polar marine phytoplankton. Org Geochem,] is available online at <http://www.sciencedirect.com/science/article/pii/S0146638017301237d> [<https://doi.org/10.1016/j.orggeochem.2017.05.007>].

Accepted Manuscript

Identification of C₂₅ highly branched isoprenoid (HBI) alkenes in diatoms of the genus *Rhizosolenia* in polar and sub-polar marine phytoplankton.

Simon T. Belt, Thomas A. Brown, Lukas Smik, Agnieszka Tatarek, Józef Wiktor, Gabriele Stowasser, Philipp Assmy, Claire S. Allen, Katrine Husum

PII: S0146-6380(17)30123-7
DOI: <http://dx.doi.org/10.1016/j.orggeochem.2017.05.007>
Reference: OG 3553

To appear in: *Organic Geochemistry*

Received Date: 9 March 2017
Revised Date: 9 May 2017
Accepted Date: 14 May 2017

Please cite this article as: Belt, S.T., Brown, T.A., Smik, L., Tatarek, A., Wiktor, J., Stowasser, G., Assmy, P., Allen, C.S., Husum, K., Identification of C₂₅ highly branched isoprenoid (HBI) alkenes in diatoms of the genus *Rhizosolenia* in polar and sub-polar marine phytoplankton., *Organic Geochemistry* (2017), doi: <http://dx.doi.org/10.1016/j.orggeochem.2017.05.007>

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.



Identification of C₂₅ highly branched isoprenoid (HBI) alkenes in diatoms of the genus *Rhizosolenia* in polar and sub-polar marine phytoplankton.

Simon T. Belt ^{a*}, Thomas A. Brown ^{a,b}, Lukas Smik ^a, Agnieszka Tatarek ^c, Józef Wiktor ^c, Gabriele Stowasser ^d, Philipp Assmy ^e, Claire S. Allen ^d, Katrine Husum ^e.

^a Biogeochemistry Research Centre, School of Geography, Earth and Environmental Sciences, University of Plymouth, Drake Circus, Plymouth, Devon PL4 8AA, UK

^b Marine Ecology and Chemistry, Scottish Association for Marine Science, Oban, Argyll, UK, PA37 1QA.

^c Institute of Oceanology Polish Academy of Sciences, Powstańców Warszawy 55, 81-712 Sopot, Poland

^d British Antarctic Survey, High Cross, Madingley Road, Cambridge, CB3 0ET, UK

^e Norwegian Polar Institute, Fram Centre, NO-9296 Tromsø, Norway

* Author for correspondence. Tel.: +44 (0)1752 584959; Fax: +44 (0)1752 584709.

E mail address: sbelt@plymouth.ac.uk (Simon Belt).

ABSTRACT

We report the identification of a range of C₂₅ highly branched isoprenoid (HBI) alkenes and certain sterols in filtered phytoplankton samples obtained from western Svalbard (Arctic) and near South Georgia (South Atlantic, sub-Antarctic) in 2016 and 2014, respectively. The C₂₅ HBIs contained 3–5 double bonds and had structures identified previously from analysis of laboratory diatom cultures. The same HBIs were also identified in individual diatom taxa isolated from the mixed assemblages and with reasonably similar distributions. Thus, C₂₅ HBIs were identified in *Rhizosolenia setigera* isolated from western Svalbard near-surface waters, while the same HBIs were also found in *R. polydactyla* f. *polydactyla* and *R. hebetata* f. *semispina* picked from seawater collected from a site in the South Atlantic. The main sterol composition was slightly different between the two locations, with cholest-5,24-dien-3 β -ol (desmosterol) identified as one of the major components in the sample from West Svalbard, consistent with the diatom assemblage being dominated by *R. setigera*. In contrast, the major sterol in the South Atlantic sample was cholest-5,22-dien-3 β -ol (22-dehydrocholesterol), likely reflecting the relatively high proportion of the genus *Pseudo-nitzschia*. For both locations, the suite of HBIs included a tri-unsaturated isomer (HBI III; 6Z-2,6,10,14-tetramethyl-9-(3'-methylpent-4-enylidene)-pentadec-6-ene), proposed in previous studies as a potential proxy measure of pelagic sea ice-edge conditions, and thus, a counterpart to the mono- and di-unsaturated HBIs IP₂₅ and IPSO₂₅, which have been used as seasonal sea ice proxies in the Arctic and Antarctic, respectively.

HBI III has been reported previously in sediments from West Svalbard and we report here its occurrence in a small number of surface sediments from the South Atlantic. For both regions, HBI III was present as one of the major HBIs in sediments, which contrasts the HBI distributions in the filtered phytoplankton samples, where HBIs with four and five double bonds were the major components. Differences in HBI distributions between phytoplankton and sediment samples may potentially be due to the presence of other (unanalysed) diatoms in the filtered water samples, seasonal/annual variability in the production of HBIs by a range of diatoms, differential degradation of HBIs between sources and sediments, or a combination of these. Interestingly, we did not detect any C₃₀ HBIs in the water samples, picked cells or sediments from either location, despite earlier reports of these lipids in laboratory cultures of *R. setigera*. This study represents the first source identification of certain C₂₅ HBI lipids under in situ pelagic conditions.

Keywords: highly branched isoprenoid; alkene; diatom; biomarker; *Rhizosolenia*

1. Introduction

C_{25} and C_{30} highly branched isoprenoid (HBI) alkenes are common components of marine and lacustrine sediments (Rowland and Robson, 1990; Belt et al., 2000; Sinninghe Damsté et al., 2004; Belt and Müller, 2013) and are generally believed to be biosynthesised by a limited number of diatom genera. To date, C_{25} HBIs (e.g. Fig. 1) have been reported in laboratory cultures of individual species of *Haslea* (Volkman et al., 1994; Belt et al., 1996; Wraige et al., 1997; Allard et al., 2001; Poulin et al., 2004), *Navicula* (Belt et al., 2001c), *Rhizosolenia* (Volkman et al., 1994; Sinninghe Damsté et al., 1999; Belt et al., 2001a, 2002; Rowland et al., 2001), *Pleurosigma* (Belt et al., 2000; 2001b; Grossi et al., 2004) and *Berkeleya* (Brown et al., 2014a). Further, under in situ environmental conditions, a small number of C_{25} HBIs have also been identified in *Pseudosolenia calcar-avis* isolated from Baltic Sea surface waters (Kaiser et al., 2016). On the other hand, apart from a limited number of reports in sediments (e.g., Prahl et al., 1980; Barrick and Hedges, 1981) and particulate organic matter (e.g., Wakeham et al., 2002; Xu and Jaffé, 2007), C_{30} HBIs have only been identified in laboratory cultures of *R. setigera* (Volkman et al., 1994; Belt et al., 2001a, 2002; Rowland et al., 2001). For both C_{25} and C_{30} HBIs, structural determinations have been achieved largely through laboratory culturing and analysis of purified apolar lipid extracts using NMR spectroscopy (e.g., Belt et al., 1996, 2000, 2001a,b,c; Sinninghe Damsté et al., 1999; Grossi et al., 2004; Brown et al., 2014a).

In recent years, the source or environmental specificity of certain C₂₅ HBI alkenes has led to their use as organic geochemical proxies for seasonal Arctic and Antarctic sea ice reconstruction (e.g., Belt et al., 2007, 2016; Massé et al., 2011; Belt and Müller, 2013). Thus, a mono-unsaturated C₂₅ HBI termed IP₂₅ (structure I; Fig. 1) has been used as a palaeo proxy for Arctic sea ice (e.g., Belt et al., 2007; Fahl and Stein., 2012; Belt and Müller, 2013; Knies et al., 2014; Müller and Stein, 2014; Stein et al., 2016), while a closely related di-unsaturated analogue (IPSO₂₅; structure II; Fig. 1) represents a likely counterpart for the Antarctic (e.g., Barbara et al., 2010, 2013; Denis et al., 2010; Massé et al., 2011; Collins et al., 2013; Etourneau et al., 2013; Belt et al., 2016). Furthermore, sources of IP₂₅ and IPSO₂₅ have been identified following isolation of individual species from mixed sea ice algal communities and analysis of their lipid content using gas chromatography–mass spectrometry (GC–MS) (Brown et al., 2014b; Belt et al., 2016). In contrast, although a tri-unsaturated C₂₅ HBI (HBI III; Fig. 1) has been suggested to be a possible proxy indicator of the retreating ice edge during spring (Collins et al., 2013; Belt et al., 2015; Smik et al., 2016a,b; Ribeiro et al., 2017), thus far, no source identification of this biomarker from such locations has been made.

In the current study, we report the occurrence of various C₂₅ HBIs and certain sterols in filtered water samples collected during (ice-free) summers from West Svalbard (Arctic) and near to South Georgia (South Atlantic, sub-Antarctic) and, in particular, we identify individual species of *Rhizosolenia* that biosynthesise HBI III.

We also believe this to be the first report of HBI source identification from in situ polar and sub-polar open water (pelagic) settings.

2. Experimental

2.1. Sample collection

Water samples were collected from western Svalbard (sample V12; 78°58.52'N; 9°21.1'E) and slightly north of South Georgia in the South Atlantic (sample E103; 53°15.56'S; 38°25.01'W) as part of the annual Kongsfjorden “Climate and Ecosystem” (Norwegian Polar Institute) and JR304 (British Antarctic Survey) cruise campaigns in 2016 and 2014, respectively (Fig. 2). All sampling was carried out in ice-free open water conditions (August and December for V12 and E103, respectively). The V12 sample was collected from a single vertical tow (0–30 m) using a plankton net (HYDRO-BIOS[®], Kiel, Germany) fitted with a 20 µm mesh. Approximately 50 ml of sampled seawater were filtered onto a 47 mm Whatman GF/F filter and kept frozen (–20 °C) prior to analysis. The E103 sample was obtained using a paired motion-compensated Bongo net (61 cm mouth diameter, 2.3 m length) equipped with solid cod-ends and 100 µm and 200 µm mesh sizes. Based on the area of the net’s mouth and the vertical sampling interval (0–200 m), we estimate the sampled volume of seawater to be ca. 58 m³. Of the 100 µm sample retrieved, ca. 2 l were filtered onto a 47 mm GF/F filter and kept frozen (–80 °C) prior to analysis. Further unfiltered aliquots of V12 and E103 (ca. 25–50 ml) were also collected and kept frozen for subsequent species identification and cell picking.

Surface sediment material from seven locations in the South Atlantic (Fig. 2) was taken from the upper 0–1 cm of archived box cores held at the British Antarctic Survey, UK.

2.2. Species identification

Centric diatoms of the genus *Rhizosolenia* have long cylindrical cells with many girdle bands and, usually, with a single, elongated, rimoportula or labiate process (spine) on each cell valve (Round et al., 1990; Scott and Thomas, 2005). Species identification using light microscopy is based, usually, on the shape of the valve and its process with associated otaria morphology (Priddle et al., 1990; Armand and Zielinski 2001). *Rhizosolenia setigera* is narrow in diameter (4–20 μm) with a long needle-like process lacking otaria. *R. hebetata* f. *semispina* is also narrow (4–25 μm), with a long tapering process, but has a small pointed otaria. In contrast, cells of *R. polydactyla* f. *polydactyla* are wider (15–105 μm) with a process that is also wider at the base, tapering to the tip, with a large otaria that tapers distally to the process.

2.3. Extraction and purification

Filtered water samples were extracted, partially purified and analysed using established methods (e.g., Belt et al., 2012, 2013). In brief, GF/F filters were saponified in methanolic KOH (ca. 4 ml $\text{H}_2\text{O}/\text{MeOH}$, 1:9; 5% KOH; 60 min, 70 °C) following addition of 9-octylheptadec-8-ene (10 ng) as internal standard to permit

quantification of HBIs. Hexane (3×2 ml) was added to the saponified solution, which was vortexed (1 min) and centrifuged (1 min; 2,000 rpm). The supernatant, containing apolar lipids, was transferred to a clean vial and dried (N_2 stream) to remove hexane and traces of $H_2O/MeOH$. The apolar fractions were re-suspended in hexane (0.5 ml) and fractionated using column chromatography (0.5 g SiO_2) to obtain HBIs (5 ml hexane) and sterols (5 ml hexane/methyl acetate (4:1, v/v)). The procedure for analysis of the picked individual diatoms was the same as for the filtered water samples, except that cells were extracted with hexane only (1 ml, ultrasonication; 5 min). Freeze-dried surface sediments (ca. 2–3 g) from the South Atlantic were extracted using dichloromethane/methanol (3×3 mL; 2:1, v/v) according to established methods (Belt et al., 2012), with the resulting lipid extracts treated as per the extracted water samples. Analysis of sediments from western Svalbard is described in Smik and Belt (2017).

2.4. Analytical methods

All lipid extracts were analysed using GC–MS in total ion current (TIC) or single ion monitoring (SIM) mode using an Agilent 7890a Series II gas chromatograph, fitted with a 30 m fused silica HP_{5ms} column (0.25 mm i.d., 0.25 μ m film) coupled to a 5975c Series Mass Selective Detector (MSD) (Belt et al., 2012). Individual HBIs were identified based on their characteristic retention indices (RI) and mass spectra (Wraige et al., 1999; Belt et al., 2000; Brown and Belt, 2016). For HBI quantification (picked cells), individual integrated peak areas for HBIs III and

IV obtained from GC–MS SIM analyses were normalised to those of the internal standard, instrumental response factors obtained from calibrations using purified standards (Belt et al., 2000, 2012) and the number of cells extracted. Since we did not have sufficient quantity and purity of all HBIs to conduct the corresponding calibrations, we took integrated peak areas of the molecular ion for each isomer and the calibrations using HBI III and IV to provide estimates of the concentrations of all other HBI components. Sterol fractions were derivatised using N,O-bis(trimethylsilyl)trifluoroacetamide (BSTFA; 50 μ l; 70 $^{\circ}$ C; 60 min) prior to analysis by GC–MS. Individual sterols were identified by comparison of the mass spectra of their TMS ethers with published data (e.g., Volkman, 1986).

3. Results

3.1. C₂₅ HBIs and sterols in phytoplankton from western Svalbard and the South Atlantic

The partially purified extracts of the filtered water samples from western Svalbard (sample V12) and South Georgia in the South Atlantic (sample E103) contained a number of C₂₅ HBIs that could be identified by comparison with previously reported GC–MS data. Thus, sample V12 (western Svalbard) contained HBIs III–VIII with VII present as the major component (Fig. 3a). HBIs III–VIII were also present in the filtered water sample from the South Atlantic (E103), with V and VII as the most abundant isomers in approximately equal amounts (Fig. 3b). In addition, relatively small amounts of HBI IX could also be identified in E103,

although its geometric isomer, X, was not detected (Fig. 3b). In contrast, C₂₅ HBIs I (IP₂₅) and II (IPSO₂₅) and C₃₀ HBIs could not be identified in water samples from either location. The main sterols in sample V12 were 24-methylcholesta-5,22-dien-3 β -ol (epi-brassicasterol), 24-methylcholesta-5,24(28)-dien-3 β -ol (24-methylenecholesterol), cholesta-5,22-dien-3 β -ol (22-dehydrocholesterol), cholesta-5,24-dien-3 β -ol (24-dehydrocholesterol or desmosterol), cholest-5-en-3 β -ol (cholesterol) and 24-methylcholest-5-en-3 β -ol (24-methylcholesterol), with desmosterol and cholesterol as the major constituents. In contrast, 22-dehydrocholesterol and cholesterol dominated the sterol composition of sample E103, with 22-dehydrocholesterol as the main component, while epi-brassicasterol, 24-methylenecholesterol and desmosterol were only present in relatively minor quantities.

3.2. C₂₅ HBIs in picked cells

The taxonomic composition of sample V12 (western Svalbard) was dominated by *Rhizosolenia setigera* (> 90% of total diatom abundance) and the same HBIs identified in the mixed microphytoplankton assemblage were also identified in the picked cells of this species, and in similar distribution, especially for the three most abundant components III, V and VII (Fig. 3a, 4a). The most abundant diatom taxa in the South Atlantic Bongo net sample (E103) were *Pseudo-nitzschia lineola* (ca. 50%) and *Trichotoxon reinboldii* (ca. 22%), with *R. polydactyla* f. *polydactyla* (ca. 11%) and *R. hebetata* f. *semispina* (3%) only present as relatively minor species. The

main HBIs in picked cells of *R. polydactyla* f. *polydactyla* and *R. hebetata* f. *semispina* were III, V and VII, although their relative concentrations were somewhat different to those of the same lipids in the total sample, with a much more even distribution in the picked cells (Figs. 3b and 4b,c). On the other hand, some other C₂₅ HBIs (e.g., IV, VI and VIII) were either absent or below the limit of detection. Unfortunately, cells of the most abundant species (*P. lineola*) were too small and difficult to remove from the glass vial walls to enable their isolation and lipid analysis. The total C₂₅ HBI concentration was estimated to be ca. 7, 3 and 2 pg/cell for *R. setigera*, *R. hebetata* f. *semispina* and *R. polydactyla* f. *polydactyla*, respectively.

3.3. C₂₅ HBIs in western Svalbard and South Atlantic surface sediments

Previously, HBI III has been reported in 27 surface sediments from western Svalbard with concentration in the range 0.27–8.78 ng/g (Smik and Belt, 2017). Here, we re-examined the GC–MS chromatograms from this previous study and identified tri-unsaturated IV as the major HBI in most cases, together with III, as reported previously, and IX as an additional minor component. In contrast, of the more unsaturated HBIs, only V could be identified, and this was only present in a few extracts and in very low relative amounts (ca. 1%; Fig. 5a). For the seven surface sediments from the South Atlantic, III was the most abundant HBI, with a concentration range of 6–250 ng/g. Similar to the western Svalbard sediments, HBI

trienes IV and IX could also be quantified, but only trace amounts of HBI V were detected (Fig. 5b).

4. Discussion

Despite the common occurrence of C_{25} HBIs in sediments (e.g., Rowland and Robson, 1990; Belt et al., 2000; Sinninghe Damsté et al., 2004; Belt and Müller, 2013), relatively few studies have reported on the presence of these lipids in their native marine or lacustrine settings, either in mixed phytoplankton assemblages or in individual taxa. Exceptionally, IP_{25} and $IPSO_{25}$ have been identified in individual and mixed assemblages of Arctic and Antarctic sea ice diatoms (Nichols et al., 1988; Belt et al., 2007, 2013, 2016; Brown et al., 2011, 2014b), di- through to penta-unsaturated C_{25} HBIs have been reported in a small number of Antarctic phytoplankton samples (Massé et al., 2011; Smik et al., 2016a), and some further di- and tri-unsaturated C_{25} HBIs were also observed in *Pseudosolenia calcar-avis* isolated from surface waters of the south-eastern Baltic Sea (Kaiser et al., 2016). P_{25} and some other HBIs have also been reported in sinking particles following the release of sympagic algae from melting sea ice in the Arctic (Brown et al., 2016; Rontani et al., 2016). As such, our identification of a range of C_{25} HBIs in phytoplankton samples from polar (western Svalbard) and sub-polar (South Atlantic) locations adds to the growing reports of these biomarkers in their source environments and we believe it to be the first example from individual taxa isolated from Arctic or South Atlantic pelagic settings.

With respect to the individual HBI-producing diatoms described in the current study, our findings represent the first report of HBIs in *R. polydactyla* f. *polydactyla* and *R. hebetata* f. *semispina*, although the occurrence of C_{25} and C_{30} HBIs within *R. setigera* is well known (Volkman et al., 1994; Sinnighe Damsté et al., 1999; 2004; Belt et al., 2001a, 2002; Rowland et al., 2001; Massé et al., 2004) and some HBIs have also been identified in *R. fallax*, *R. shrubshrolei* and *R. pungens* (Sinnighe Damsté et al., 2004). The absence of any C_{30} HBIs is also intriguing given their biosynthesis by *R. setigera* in most laboratory cultures (Volkman et al., 1994; Belt et al., 2001a, 2002; Rowland et al., 2001). On the other hand, C_{30} HBIs were also absent in cultures of *R. setigera* isolated from the east coast of the USA (Sinnighe Damsté et al., 1999), although this strain was additionally unusual in that it produced only one (penta-unsaturated) C_{25} HBI and with a double bond at C5/6 compared to C7/20, which is a more common characteristic of C_{25} and C_{30} HBIs in other strains of *R. setigera* (Belt et al., 2001a, 2002; Rowland et al., 2001). However, even within the C_{30} HBI-producing strains, the presence and distribution of the C_{25} counterparts exhibit notable differences. For example, Volkman et al. (1994) first reported the occurrence of several C_{30} HBIs (but no C_{25} HBIs) in an Australian strain (CS-62) of *R. setigera*, and Belt et al. (2001a) reported similar findings for a further strain (Nantes 99) isolated from northern France. In contrast, Rowland et al. (2001) detected both C_{25} (including III–VI identified here) and C_{30} HBIs in an Australian strain of *R. setigera* (CS 389/A), while Belt et al. (2002) showed subsequently that their distribution was strongly

influenced by life cycle characteristics, with the biosynthesis of C_{25} HBIs, in particular, being stimulated during the sexual reproduction or auxosporulation stage. In any case, the absence of C_{30} HBIs in our mixed phytoplankton and individual *Rhizosolenia* diatoms isolated from natural surface waters may potentially explain the relatively small number of reports of these biomarkers in marine sediments, at least compared to their C_{25} pseudo-homologues (Rowland and Robson, 1990; Belt et al., 2000; Sinninghe Damsté et al., 2004; Belt and Müller, 2013). On the other hand, the identification of desmosterol as the major sterol in the *R. setigera*-rich V12 sample is consistent with previous findings from laboratory cultures (Barrett et al., 1995; Massé et al., 2004; Rampen et al., 2010). Similarly, the presence of 22-dehydrocholesterol as the major sterol in sample E103 from the South Atlantic is consistent with the occurrence of *Pseudo-nitzschia lineola* as the most abundant diatom. Thus, although we are not aware of any investigations into the sterol content of *P. lineola* in culture, Rampen et al. (2010) identified 22-dehydrocholesterol as the major sterol in *P. seriata*.

In addition to the variability in HBI composition within *Rhizosolenia* diatoms, the type, concentration and distribution of individual isomers identified in V12, E103 and picked cells from both of these mixed algal assemblages, exhibit some parallels with HBI content in other diatoms, even in those of diverse (phylogenetically) genera. For example, the co-occurrence of III–VIII found here in centric *Rhizosolenia* diatoms has been reported previously in laboratory cultures of the pennate diatom *Pleurosigma intermedium* (Belt et al., 2000), which is also

capable of biosynthesising IX (Brown and Belt, 2016). Furthermore, our estimates of cellular (total) HBI concentrations (ca. 2–6 pg/cell) are typical of those reported previously in laboratory cultures of HBI-producing diatoms (Volkman et al., 1994; Rowland et al., 2001; Massé et al., 2004; Belt et al., 2013; Brown et al., 2014a; Kaiser et al., 2016) and individual species isolated from natural ice-algal assemblages (Brown et al., 2014b; Belt et al., 2016).

For all three *Rhizosolenia* species, we note, in particular, the presence of a tri-unsaturated C_{25} HBI (HBI III) that has been proposed as a potential proxy for ice-edge pelagic conditions in both the Arctic and the Antarctic (Collins et al., 2013; Belt et al., 2015; Smik et al., 2016a,b; Ribeiro et al., 2017). Given the near-ubiquity of *Rhizosolenia* spp. in marine phytoplankton worldwide, including the Arctic and Antarctic (Priddle and Fryxell, 1985; Priddle et al., 1990; Scott and Thomas, 2005), it seems likely that the *Rhizosolenia* species identified here contribute to the sedimentary budget of HBI III in certain polar and sub-polar environments. Previously, HBI III has been reported in surface and down-core sediments from western Svalbard (Cabedo-Sanz and Belt, 2016; Smik et al., 2017) and we also identified it in each of the surface sediments from the South Atlantic as part of the current study, so a combination of our new and previous findings suggest that *Rhizosolenia* spp. are likely sources. However, since only a single sample was collected from each region, and these were both from ice-free surface waters during spring/summer months, the results from the current study do not really add to the evidence described previously for the use of HBI III as a proxy for ice-edge

conditions in the Arctic and the Antarctic (Collins et al., 2013; Belt et al., 2015; Smik et al., 2016a,b; Ribeiro et al., 2017). Further, our study does not reveal whether HBI III (or other HBIs) might be biosynthesised by other diatoms in these regions that bloom during other intervals. An examination of a greater number of diatom species is therefore required before the contribution from *Rhizosolenia* spp. in polar and sub-polar environments can be fully evaluated.

For both western Svalbard and the South Atlantic study regions, the sedimentary HBI distributions differ, however, from those found in the filtered water samples or individual diatom taxa. Specifically, while the tetra- and penta-unsaturated HBIs V and VII were present as the major components in the samples of filtered water and picked diatoms from both regions (Fig. 3), HBI trienes (III, IV and IX) were the most significant constituents of the surface sediments, with only V as the other quantifiable HBI, and in very low amounts (Fig. 5). We offer three possible explanations for these differences.

First, the snapshot nature of our phytoplankton sampling likely limits the extent to which the corresponding HBI distributions parallel those that reflect accumulation over seasonal or annual timeframes that are pertinent to sediments. As described earlier, there may be further diatoms in these regions that biosynthesise HBIs during different seasons, such that sedimentary distributions may better reflect the collective contribution resulting from seasonal species succession. Thus, additional phytoplanktonic sources of HBIs such as III, IV and IX would likely result in their increased accumulation, relative to HBIs V–VIII, in

sediments. To date, the only other known sources of HBIs III, IV and IX are diatoms belonging to the genus *Pleurosigma* (Belt et al., 2000; Brown and Belt, 2016), but *Pleurosigma* spp. were either absent or only present in extremely low abundances in our samples. However, this does not discount the possibility of HBI production by *Pleurosigma* spp. or other diatoms during different seasons, or by unpicked species in the current samples. Indeed, we note that the distributions of HBIs III, V and VII in *R. polydactyla* f. *polydactyla* and *R. hebetata* f. *semispina* (Fig. 4b,c) were slightly different from that in the mixed phytoplankton sample from which they were picked (E103; Fig. 3b), indicating the likely occurrence of additional HBI-producers in the latter. Further, and in contrast to the HBI distributions in the filtered phytoplankton and picked cells from sample V12, the identification of IX and the increased relative abundance of IV compared to III in sediments from western Svalbard (Fig. 5a), indicate that species other than *R. setigera* potentially contribute to the HBI sedimentary budget in this region. On the other hand, the contrasting outcomes between phytoplankton and sedimentary analyses may simply reflect the variability in HBI distribution observed previously in *Rhizosolenia* spp. (Volkman et al., 1994; Sinninghe Damsté et al., 1999; Belt et al., 2001a, 2002; Rowland et al., 2001), with sediment composition indicative of a temporal average of any shorter-term HBI variability within this genus.

Second, the likely increased degradation rates of more unsaturated HBIs such as V–VIII compared to those of HBI trienes (i.e. III, IV and IX) potentially leads to the latter becoming relatively enhanced in sediments. Indeed, although a

direct comparison of the reactivity of HBIs III–IX under environmental conditions has not yet been carried out, in laboratory studies a general increase in reactivity towards photo- and autoxidation processes has been reported for some HBIs containing a larger number of double bonds (Rontani et al., 2011, 2014).

Third, some additional (smaller) HBI-producing diatoms may not have been obtained during water sample collection in the South Atlantic, especially, due to the increased mesh size of the Bongo net employed (100 μm). In any case, the extent to which *Pleurosigma*, or other diatom genera, are additional contributors to the sedimentary budget of HBI III (or other HBIs) will require analysis of a larger number of phytoplankton samples with variable diatom composition. For now, although we were not able to isolate individual cells of the abundant (ca. 50%) *Pseudonitzschia lineola* from sample E103, we note that *P. seriata* has been shown previously not to produce HBIs in culture (Sinninghe Damsté et al., 2004).

5. Conclusions

A number of C_{25} HBI alkenes have been identified in natural phytoplankton populations obtained from West Svalbard in the Arctic and north of South Georgia in the South Atlantic (sub-Antarctic), including a tri-unsaturated isomer (HBI III) proposed previously as a potential proxy for seasonal ice-edge conditions in polar and sub-polar settings. From the same samples, picked diatoms belonging to the genus *Rhizosolenia* contained similar distributions of HBIs to those of the mixed phytoplankton assemblages, although they exhibited clear differences to those in

surface sediments from each region and also those reported previously in laboratory cultures of *R. setigera*, with the absence of any C₃₀ HBIs being particularly noteworthy. In contrast, the identification of desmosterol as the major sterol in the sample from West Svalbard, containing > 90% *R. setigera*, is consistent with previous investigations into diatom sterol composition. In the future, it will be important to determine whether any other diatoms are capable of producing C₂₅ HBIs (especially HBI III) in other polar and sub-polar pelagic settings, and to investigate whether there are any specific environmental controls (e.g., season) over HBI production in order that their potential as palaeoenvironmental proxies can be better understood. Such investigations are currently underway in our laboratories.

Acknowledgments

This work was supported by the University of Plymouth and a Research Project Grant awarded by the Leverhulme Trust. The western Svalbard material was collected as part of the long-term monitoring of Kongsfjorden and neighbouring shelf by the Norwegian Polar Institute. Core sediments presented here from the South Atlantic were collected aboard the RRS James Clark Ross during cruise JR257 in 2012. We thank the Captain and crew of the RRS James Clark Ross and scientific party of JR257 and JR304 for their support. Finally, we thank the supportive and useful comments from two anonymous reviewers and the Associate Editor (Dr Mark Yunker), which helped to improve the clarity of this manuscript.

Associate Editor–Mark Yunker

References

- Allard, W. G., Belt, S.T., Massé, G., Naumann, R., Robert, J.-M., Rowland, S.J.,
2001. Tetra-unsaturated sesterterpenoids (Haslenes) from *Haslea ostrearia*
and related species. *Phytochemistry* 56, 795–800.
- Armand, L.K., Zielinski, U. 2001. Diatom species of the genus *Rhizosolenia* from
Southern Ocean sediments: Distribution and taxonomic notes. *Diatom
Research* 16, 259–294.
- Barbara, L., Crosta, X., Massé, G., Ther, O., 2010. Deglacial environments in eastern
Prydz Bay, East Antarctic. *Quaternary Science Reviews* 29, 2731–2740.
- Barbara, L., Crosta, X., Schmidt, S., Massé, G., 2013. Diatoms and biomarkers
evidence for major changes in sea ice conditions prior the instrumental period
in Antarctic Peninsula. *Quaternary Science Reviews* 79, 99–110.
- Barrett, S.M., Volkman, J.K., Dunstan, G.A., 1995. Sterols of 14 species of marine
diatoms (Bacillariophyta). *Journal of Phycology* 31, 360–369.
- Barrick, R.C., Hedges, J.I., 1981. Hydrocarbon geochemistry of the Puget Sound
region – II. Sedimentary diterpenoid, steroid and triterpenoid hydrocarbons.
Geochimica et Cosmochimica Acta 45, 381–392.
- Belt, S.T., Cooke, D.A., Robert, J.-M., Rowland, S.J., 1996. Structural characterisation
of widespread polyunsaturated isoprenoid biomarkers: A C₂₅ triene, tetraene

- and pentaene from the diatom *Haslea ostrearia* Simonsen. *Tetrahedron Letters* 37, 4755–4758.
- Belt, S.T., Allard, W.G., Massé, G., Robert, J.-M., Rowland, S.J., 2000. Highly branched isoprenoids (HBIs): identification of the most common and abundant sedimentary isomers. *Geochimica et Cosmochimica Acta* 64, 3839–3851.
- Belt, S.T., Allard, W.G., Massé, G., Robert, J.-M., Rowland, S.J., 2001a. Structural characterisation of C₃₀ highly branched isoprenoid alkenes (rhizenes) in the marine diatom *Rhizosolenia setigera*. *Tetrahedron Letters* 42, 5583–5585.
- Belt, S.T., Massé, G., Allard, W.G., Robert, J.-M., Rowland, S.J., 2001b. C₂₅ highly branched isoprenoid alkenes in planktonic diatoms of the *Pleurosigma* genus. *Organic Geochemistry* 32, 1271–1275.
- Belt, S.T., Massé, G., Allard, W.G., Robert, J.-M., Rowland, S.J., 2001c. Identification of a C₂₅ highly branched isoprenoid triene in the freshwater diatom *Navicula sclesvicensis*. *Organic Geochemistry* 32, 1169–1172.
- Belt, S.T., Massé, G., Allard, W.G., Robert, J.-M., Rowland, S.J., 2002. Effects of auxosporulation on distributions of C₂₅ and C₃₀ isoprenoid alkenes in *Rhizosolenia setigera*. *Phytochemistry* 59, 141–149
- Belt, S.T., Massé, G., Rowland, S.J., Poulin, M., Michel, C., LeBlanc, B., 2007. A novel chemical fossil of palaeo sea ice: IP₂₅. *Organic Geochemistry* 38, 16–27.

- Belt, S.T., Brown, T.A., Navarro Rodriguez, A., Cabedo Sanz, P., Tonkin, A., Ingle, R., 2012. A reproducible method for the extraction, identification and quantification of the Arctic sea ice proxy IP₂₅ from marine sediments. *Analytical Methods* 4, 705–713.
- Belt, S.T., Brown, T.A., Ringrose, A.E., Cabedo-Sanz, P., Mundy, C.J., Gosselin, M., Poulin, M., 2013. Quantitative measurements of the sea ice diatom biomarker IP₂₅ and sterols in Arctic sea ice and underlying sediments: Further considerations for palaeo sea ice reconstruction. *Organic Geochemistry* 62, 33–45.
- Belt, S.T., Müller, J., 2013. The Arctic sea ice biomarker IP₂₅: a review of current understanding, recommendations for future research and applications in palaeo sea ice reconstructions. *Quaternary Science Reviews* 79, 9–25.
- Belt, S.T., Cabedo-Sanz, P., Smik, L., Navarro-Rodriguez, A., Berben, S.M.P., Knies, J., Husum, K., 2015. Identification of paleo Arctic winter sea ice limits and the marginal ice zone: optimised biomarker-based reconstructions of late Quaternary Arctic sea ice. *Earth and Planetary Science Letters* 431, 127–139.
- Belt, S.T., Smik, L., Brown, T.A., Kim, J.-H., Rowland, S.J., Allen, C.S., Gal, J.-K., Shin, K.-H., Lee, J.I., Taylor, K.W.R., 2016. Source identification and distribution reveals the potential of the geochemical Antarctic sea ice proxy IPSO₂₅. *Nature Communications* 7, 12655.
- Brown, T.A., Belt, S.T., Mundy, C., Philippe, B., Massé, G., Poulin, M., Gosselin, M., 2011. Temporal and vertical variations of lipid biomarkers during a bottom

- ice diatom bloom in the Canadian Beaufort Sea: further evidence for the use of the IP₂₅ biomarker as a proxy for spring Arctic sea ice. *Polar Biology* 34, 1857–1868.
- Brown, T.A., Belt, S.T., Cabedo-Sanz, P., 2014a. Identification of a novel di-unsaturated C₂₅ highly branched isoprenoid in the marine tube-dwelling diatom *Berkeleya rutilans*. *Environmental Chemistry Letters*, 12, 455–460.
- Brown, T.A., Belt, S.T., Tatarek, A., Mundy, C.J., 2014b. Source identification of the Arctic sea ice proxy IP₂₅. *Nature Communications* 5, 4197.
- Brown, T.A., Belt, S.T., Gosselin, M., Levasseur, M., Poulin, M., Mundy, C.J., 2016. Quantitative estimates of sinking sea ice particulate organic carbon based on the biomarker IP₂₅. *Marine Ecology Progress Series* 546, 17–29.
- Brown, T.A., Belt, S.T., 2016. Novel tri- and tetra-unsaturated highly branched isoprenoid (HBI) alkenes from the marine diatom *Pleurosigma intermedium*. *Organic Geochemistry* 91, 120–122.
- Cabedo-Sanz, P., Belt, S.T., 2016. Seasonal sea ice variability in eastern Fram Strait over the last 2,000 years. *Arktos* 2, 22.
- Collins, L.G., Allen, C.S., Pike, J., Hodgson, D.A., Weckström, K., Massé, G. 2013. Evaluating highly branched isoprenoid (HBI) biomarkers as a novel Antarctic sea-ice proxy in deep ocean glacial age sediments. *Quaternary Science Reviews* 79, 87–98.
- Denis, D., Crosta, X., Barbara, L., Massé, G., Renssen, H., Ther, O., Giraudeau, J., 2010. Sea ice and wind variability during the Holocene in East Antarctica:

- insight on middle-high latitude coupling. *Quaternary Science Reviews* 29, 3709–3719.
- Etourneau, J., Collins, L.G., Willmott, V., Kim, J.-H., Barbara, L., Leventer, A., Schouten S., Sinninghe Damste, J.S., Bianchini, A., Klien, V., Crosta, X., Massé, G. 2013. Holocene climate variations in the western Antarctic Peninsula: evidence for sea ice extent predominantly controlled by changes in insolation and ENSO variability. *Climate of the Past* 9, 1431–1446.
- Fahl, K., Stein, R., 2012. Modern seasonal variability and deglacial/Holocene change of central Arctic Ocean sea-ice cover: new insights from biomarker proxy records. *Earth and Planetary Science Letters* 351–352, 123–133.
- Grossi, V., Beker, B., Genevasen, J.A.J., Schouten, S., Raphel, D., Fontaine, M.-F., Sinninghe Damsté, J.S., 2004. C₂₅ highly branched isoprenoid alkenes from the marine benthic diatom *Pleurosigma strigosum*. *Phytochemistry* 65, 3049–3055.
- Kaiser, J., Belt, S.T., Tomczak, T., Brown, T.A., Wasmund, N., Arz, H.W., 2016. C₂₅ highly branched isoprenoid alkenes in the Baltic Sea produced by the marine planktonic diatom *Pseudosolenia calcar-avis*. *Organic Geochemistry* 93, 51–58.
- Knies, J., Cabedo-Sanz, P., Belt, S.T., Baranwal, S., Fietz, S., Rosell-Melé, A., 2014. The emergence of modern sea ice cover in the Arctic Ocean. *Nature Communications* 5, 5608.
- Massé, G., Belt, S.T., Rowland, S.J., Rohmer, M., 2004. Isoprenoid biosynthesis in the diatoms *Rhizosolenia setigera* (Brightwell) and *Haslea ostrearia*

- (Simonsen). Proceedings of the National Academy of Sciences of the USA 101, 4413–4418.
- Massé, G., Belt, S.T., Crosta, X., Schmidt, S., Snape, I., Thomas, D.N., Rowland, S.J., 2011. Highly branched isoprenoids as proxies for variable sea ice conditions in the Southern Ocean. *Antarctic Science* 23, 487–498.
- Müller, J., Stein, R., 2014. High-resolution record of late glacial sea ice changes in Fram Strait corroborates ice-ocean interactions during abrupt climate shifts. *Earth and Planetary Science Letters* 403, 446–455.
- Nichols, P.D., Volkman, J.K., Palmisano, A.C., Smith, G.A., White, D.C., 1988. Occurrence of an isoprenoid C₂₅ diunsaturated alkene and high neutral lipid content in Antarctic sea-ice diatom communities. *Journal of Phycology* 24, 90–96.
- Poulin, M., Massé, G., Belt, S.T., Delavault, P., Rousseau, F., Robert, J.-M. and Rowland, S.J., 2004. Morphological, biochemical and molecular evidence for the transfer of *Gyrosigma nipkowii* Meister to the genus *Haslea* (Bacillariophyta). *European Journal of Phycology* 39, 181–195.
- Prahl, F. G., Bennet, J.T., Carpenter, R., 1980. The early diagenesis of aliphatic hydrocarbons and organic matter in sedimentary particles from Dabob Bay, Washington. *Geochimica Cosmochimica Acta* 44, 1967–76.
- Priddle J., Fryxell G., 1985. Handbook of the common plankton diatoms of the Southern Ocean: Centrales except the genus *Thalassiosira*, British Antarctic Survey, Cambridge, pp. 73–95.

- Priddle J., Jordan R.W., Medlin L.K., 1990. Family Rhizosoleniaceae. In: Medlin L. K., Priddle J. (Eds.), *Polar Marine Diatoms*. British Antarctic Survey, Cambridge. pp. 115–127.
- Rampen, S.W., Abbas, B.A., Schouten, S., Sinninghe Damsté, J.S.D., 2010. A comprehensive study of sterols in marine diatoms (Bacillariophyta): Implications for their use as tracers for diatom productivity. *Limnology and Oceanography* 55, 91–105.
- Ribeiro, S., Sejr, M.K., Limoges, A., Heikkilä, M., Andersen, T.J., Tallberg, P., Weckström, K., Husum, K., Forwick, M., Dalsgaard, T., Massé, G., Seidenkrantz, M.-S., Rysgaard, S., 2017. Sea ice and primary production proxies in surface sediments from a High Arctic Greenland fjord: Spatial distribution and implications for palaeoenvironmental studies. *Ambio* 46 (Suppl. 1) S106–S118. DOI 10.1007/s13280-016-0894-2.
- Rontani, J.-F., Belt, S.T., Vaultier, F., Brown, T.A., 2011. Visible light induced photo-oxidation of highly branched isoprenoid (HBI) alkenes: Significant dependence on the number and nature of double bonds. *Organic Geochemistry* 42, 812–822.
- Rontani, J.-F., Belt, S.T., Vaultier, F., Brown, T.A., Massé, G., 2014. Autoxidation and photooxidation of highly branched isoprenoid (HBI) alkenes: a combined kinetic and mechanistic study. *Lipids* 49, 481–494.
- Rontani, J.-F., Belt, S.T., Brown, T.A., Amiraux, R., Gosselin, M., Vaultier, F., Mundy, C.J., 2016. Monitoring abiotic degradation in sinking versus

- suspended Arctic sea ice algae during a spring ice melt waters using specific lipid oxidation tracers. *Organic Geochemistry* 98, 82–97.
- Round, F.E., Crawford, R.M., Mann, D.G. 1990. *The Diatoms; Biology and morphology of the genera*. Cambridge University Press, Cambridge.
- Rowland, S.J., Robson, J.N., 1990. The widespread occurrence of highly branched acyclic C₂₀, C₂₅ and C₃₀ hydrocarbons in recent sediments and biota-A review. *Marine Environmental Research* 30, 191–216.
- Rowland, S.J., Allard, W.G., Belt, S.T., Massé, G., Robert, J.-M., Blackburn, S., Frampton, D., Revill, A.T., Volkman, J.K., 2001. Factors influencing the distributions of polyunsaturated terpenoids in the diatom, *Rhizosolenia setigera*. *Phytochemistry* 58, 717–728.
- Scott, F.J., Thomas, D.P., 2005. Diatoms. In: Scott, F.J., Marchant, H.J. (Eds.), *Antarctic Marine Protists*. Australian Biological Resources Study; Australian Antarctic Division, Canberra and Hobart, Australia, pp. 13–201.
- Sinninghe Damsté, J.S., Schouten, S., Rijpstra, W.I.C., Hopmans, E.C., Peletier, H., Gieskes, W.W.C., Genevasen, J.A.J., 1999. Structural identification of the C₂₅ highly branched isoprenoid pentaene in the marine diatom *Rhizosolenia setigera*. *Organic Geochemistry* 30, 1581–1583.
- Sinninghe Damsté, J.S., Muijzer, G., Abbas, B., Rampen, S.W., Massé, G., Allard, W.G., Belt, S.T., Robert, J.-M., Rowland, S.J., Moldowan, J.M., Barbanti, S.M., Fago, F.J., Denisevich, P., Dahl, J., Trindade, L.A.F., Schouten, S., 2004. The rise of the rhizosolenid diatoms. *Science* 304, 584–587.

- Smik, L., Belt, S.T., Lieser, J., Armand, L.K., Leventer, A., 2016a. Variations in algal lipid distributions in seasonally sea-ice covered surface waters from East Antarctica: further insights for biomarker-based paleo sea-ice reconstruction. *Organic Geochemistry* 95, 71–80.
- Smik, L., Cabedo-Sanz, P., Belt, S.T., 2016b. Semi-quantitative estimates of paleo Arctic sea ice concentration based on source-specific highly branched isoprenoid alkenes: a further development of the PIP_{25} index. *Organic Geochemistry* 92, 63–69.
- Smik, L., Belt, S.T., 2017. Distributions of the Arctic sea ice biomarker proxy IP_{25} and two phytoplanktonic biomarkers in surface sediments from West Svalbard. *Organic Geochemistry* (in press).
<http://dx.doi.org/10.1016/j.orggeochem.2017.01.005>.
- Stein, R., Fahl, K., Schreck, M., Knorr, G., Niessen, F., Forwick, M., Gebhardt, C., Jensen, L., Kaminski, M., Kopf, A., Matthiessen, J., Jokat, L., Lohmann, G., 2016. Evidence for ice-free summers in the late Miocene central Arctic Ocean. *Nature Communications* 7, 11148.
- Volkman, J.K., 1986. A review of sterol markers for marine and terrigenous organic matter. *Organic Geochemistry* 9, 83–99
- Volkman, J.K., Barrett, S.M., Dunstan, G.A., 1994. C_{25} and C_{30} highly branched isoprenoid alkenes in laboratory cultures of two marine diatoms. *Organic Geochemistry* 21, 407–414.

- Wakeham, S.G., Peterson, M.L., Hedges, J.I., Lee, C., 2002. Lipid biomarker fluxes in the Arabian Sea, with a comparison to the equatorial Pacific Ocean. *Deep-Sea Research II* 49, 2265–2301.
- Wraige, E.J., Belt, S.T., Lewis, C.A., Cooke, D.A., Robert, J.-M., Massé, G., Rowland, S.J., 1997. Variations in structures and distributions of C₂₅ highly branched isoprenoid (HBI) alkenes in cultures of the diatom, *Haslea ostrearia* (Simonsen). *Organic Geochemistry* 27, 497–505.
- Wraige, E.J., Johns, L., Belt, S.T., Massé, G., Robert, J. -M., Rowland, S.J., 1999. Highly branched C₂₅ isoprenoids in axenic cultures of *Haslea ostrearia*. *Phytochemistry* 51, 69–73.
- Xu, Y., Jaffé, R., 2007. Lipid biomarkers in suspended particles from a subtropical estuary: Assessment of seasonal changes in sources and transport of organic matter *Marine Environmental Research* 64, 666–678.

Figures and Tables

Fig. 1. Structures of C_{25} HBI alkenes described in this study.

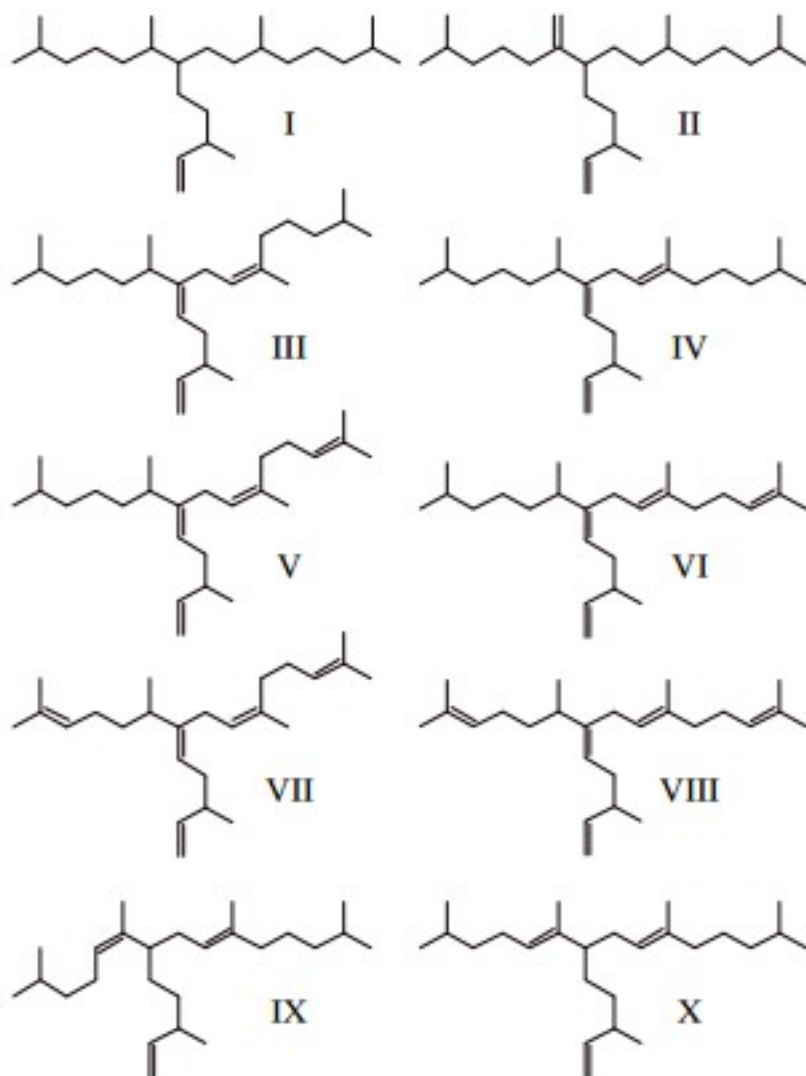
Fig. 2. Map of sampling regions: (a) western Svalbard; (b) South Atlantic. The water sample locations are indicated with a red dot. The locations of surface sediments analysed for HBIs in previous studies (Smik and Belt, 2017) and the current investigation are indicated mainly with black dots. Locations indicated by yellow dots represent the surface sediments for which partial GC–MS data are shown in Fig. 5.

Fig. 3. Partial GC–MS chromatograms (SIM mode) of extracted water samples: (a) V12; (b) E103. In each case, the selected ion corresponds to the molecular ion of C_{25} HBIs with different degrees of unsaturation (m/z 346: $C_{25:3}$; m/z 344: $C_{25:4}$; m/z 342: $C_{25:5}$). Labelled peaks correspond to the structures shown in Fig. 1. Values in parentheses refer to the % contribution of the selected HBI to the total HBI content.

Fig. 4. Partial GC–MS chromatograms of partially purified hexane extracts of picked cells of different diatoms: (a) *R. setigera*; (b) *R. polydactyla* f. *polydactyla*; (c) *R. hebetata* f. *semispina*. In each case, the selected ion corresponds to the molecular ion of C_{25} HBIs with different degrees of unsaturation as per Fig. 3. Labelled peaks

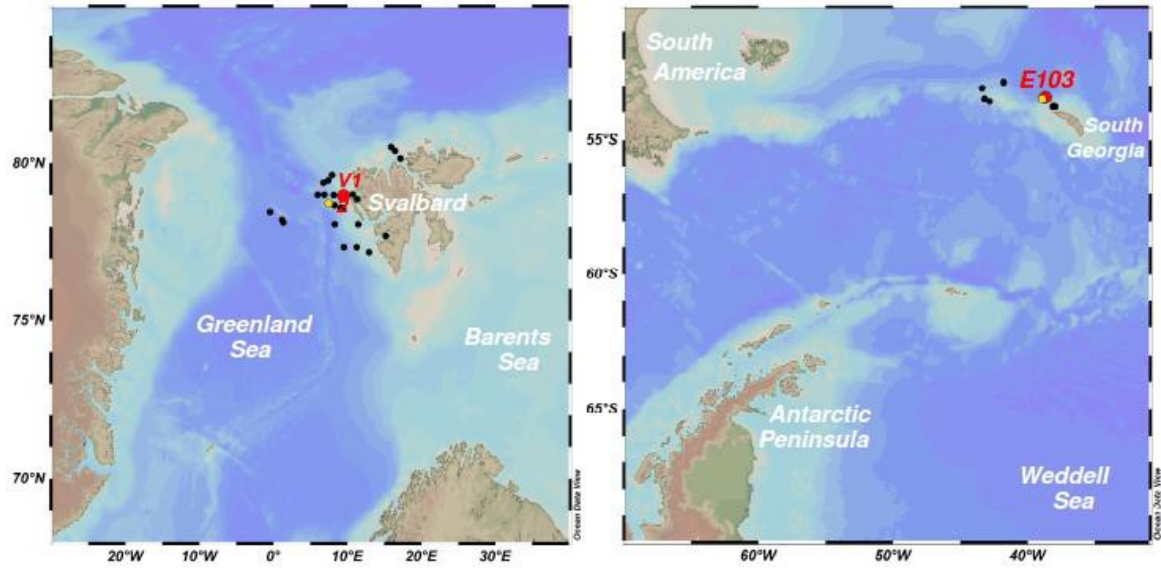
correspond to the structures shown in Fig. 1. Values in parentheses refer to the % contribution of the selected HBI to the total HBI content.

Fig. 5. Partial GC–MS chromatograms of partially purified hexane extracts of selected surface sediments: (a) western Svalbard (V12); (b) South Atlantic (E103). In each case, the selected ion corresponds to the molecular ion of C₂₅ HBIs with different degrees of unsaturation as per Fig. 3. Labelled peaks correspond to the structures shown in Fig. 1. Values in parentheses refer to the % contribution of the selected HBI to the total HBI content. For consistency with Fig. 3 and 4, the retention time of HBI VII is indicated by a dashed vertical line, although it was below the limit of detection for all sediments.



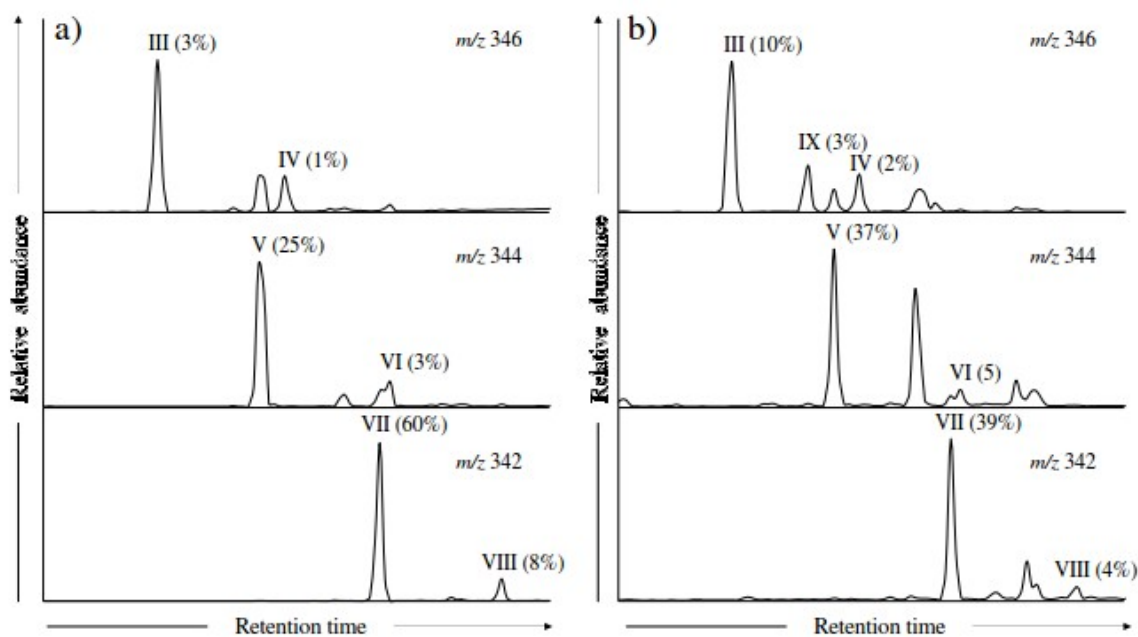
ACCEPTED

MANUSCRIPT



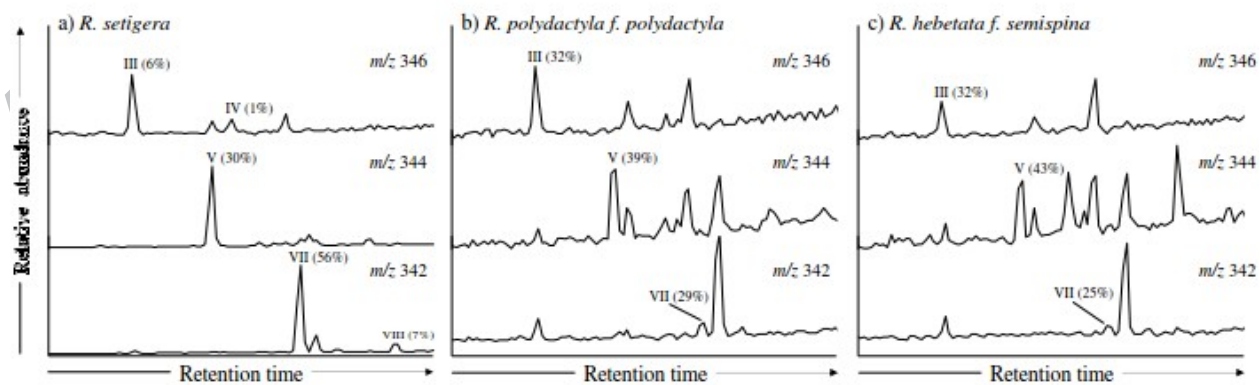
1

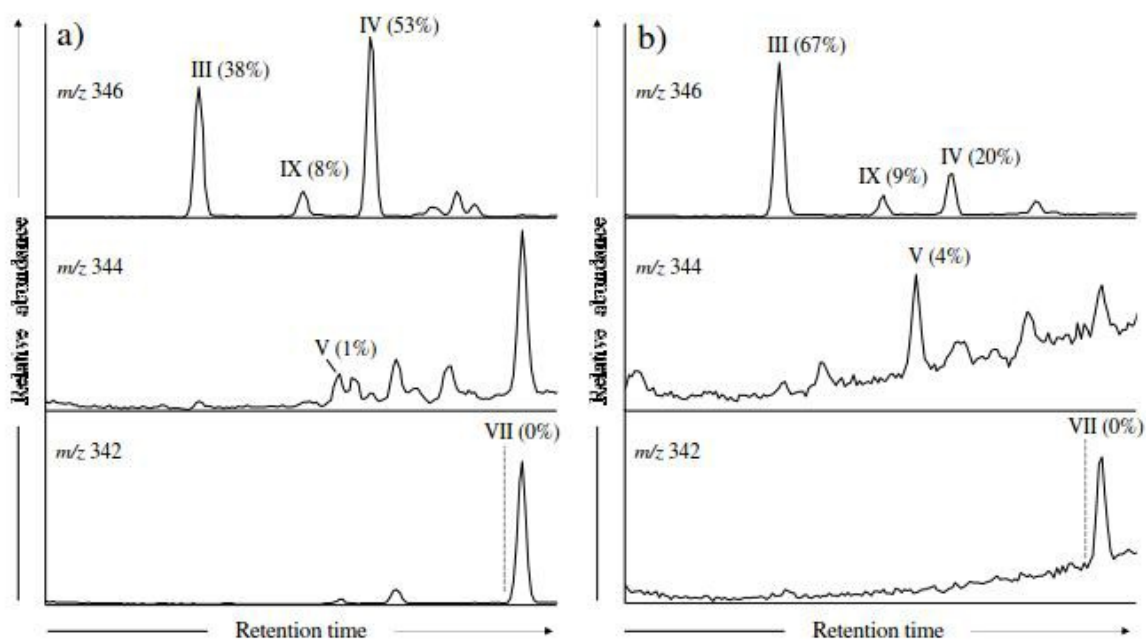
ACCEPTED



1

DTE





1

Highlights

C₂₅ HBIs identified in phytoplankton from western Svalbard and the South Atlantic

Sources of C₂₅ HBIs identified as three species of *Rhizosolenia*

HBIs include HBI III proposed previously as a possible sea ice-edge proxy

Phytoplankton sterol content consistent with laboratory cultures of major taxa

ACCEPTED MANUSCRIPT